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(71) **Applicant (for all designated States except US):** **WYETH**
[US/US]; 5 Giralda Farms, Madison, NJ 07940 (US).

(72) **Inventors; and**

(75) **Inventors/Applicants (for US only):** **SINACORE, Martin, S.** [US/US]; 32 Reservation Road, Andover, MA 01810 (US). **LEONARD, Mark** [IE/US]; 17 Cutler Lane, Manchester, NH 03104 (US). **LAKEN, Haley, A.** [US/US]; 38 Lawrence Street, Pepperell, MA 01463 (US). **ROUSE, Jason** [US/US]; 5 Lantern Lane, Londonderry, NH 03053 (US).

(74) **Agents:** **MYERS, Louis et al.; Fish & Richardson P.C.,**
P.O. Box 1022, Minneapolis, Minnesota 55440-1022 (US).

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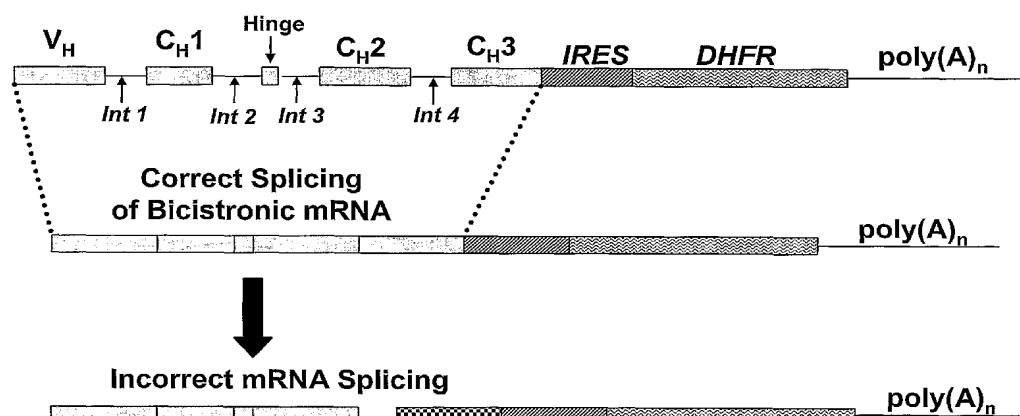
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(54) Title: METHODS AND COMPOSITIONS FOR IMPROVING RECOMBINANT PROTEIN PRODUCTION



(57) Abstract: Nucleic acid molecules modified to enhance recombinant protein, e.g., antibody, expression and/or reduce or eliminate mis-spliced and/or intron read-through (IRT) by-products are disclosed. The invention also provides methods for producing proteins devoid of mis-spliced and/or intron read-through by-products by the use of such vectors in host cells under cell culture conditions suitable for recombinant protein expression.

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METHODS AND COMPOSITIONS FOR IMPROVING RECOMBINANT PROTEIN PRODUCTION

Cross-Reference to Related Application

This application claims priority to U.S. Patent Application Serial No.

- 10 60/616,474, filed on October 5, 2004, the entire contents of which are hereby incorporated by reference.

Background of the Invention

Expression vectors for the production of recombinant proteins have existed since at least the mid 1980s. Typically, vector-based strategies for recombinant protein expression have largely been employed in basic research and for small-scale experimentation where the absolute purity of a protein preparation is not critical. In contrast, when recombinant proteins are used for therapeutic applications, even minor contaminants, for example, the presence of mis-spliced or intron read-through by-products can diminish the activity and yield of the resultant therapeutic proteins.

20 Administration of therapeutic proteins having mis-spliced or read-through protein sequences to patients may increase the possibility of undesirable side effects.

Such by-products are also troublesome for manufacturing. The presence of by-products can compromise the purification process because such by-products are typically similar to the desired proteins in terms of size, affinity, or bioactivity. Still further, it has been observed that scaling up protein expression using recombinant host cells typically results in increasing amounts of by-products as compared to the desired product, particularly if the cells are cultured under less than optimal cell culture conditions. Such sub-optimal cell culture conditions frequently occur in large scale protein production, for example, at the end of a biofermenter run or when, for other reasons, where the health of the large scale culture deteriorates.

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Accordingly, there exists a need for methods for improving recombinant protein production, particularly, for the large-scale production of therapeutic proteins.

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Summary of the Invention

The present invention provides methods and compositions for improving recombinant protein or peptide expression and/or production. In one embodiment,

5 nucleic acid molecules are provided that are modified to reduce or eliminate mis-spliced and/or intron read-through by-products, and/or to enhance recombinant protein expression. In certain embodiments, the nucleic acids encode recombinant antibodies (also referred to herein as immunoglobulins), or fragments thereof. The invention further includes vectors (e.g., expression vectors) modified to reduce or
10 eliminate mis-spliced and/or intron read-through by-products and/or to enhance recombinant protein expression; host cells, e.g., mammalian host cells, including such nucleic acid molecules and vectors; and methods for culturing such cells to produce the recombinant proteins or peptides, e.g., in large-scale. Compositions, e.g., pharmaceutical compositions, of recombinant proteins or peptides, e.g., antibodies,
15 substantially free of mis-spliced and/or intron read-through products, are also disclosed. These compositions are suitable for therapeutic use, including, for example, the treatment of neurodegenerative and malignant disorders.

Accordingly, in one aspect, the invention features a nucleic acid molecule (e.g., a modified or recombinant nucleic acid molecule) that includes a nucleotide sequence having one or more intron and exon sequences, wherein at least one intron sequence has been modified compared to the naturally-occurring sequence to enhance protein expression and/or reduce or eliminate mis-spliced or intron read-through (IRT) by-product(s). In one embodiment, the nucleic acid molecule directs enhanced expression and/or reduces or eliminates intron read-through (IRT) by-product(s) of a desired protein or peptide, for example, an antibody or a fragment thereof (e.g., an immunoglobulin heavy chain) relative to a naturally occurring sequence (e.g., a genomic sequence). The protein or peptide can be of mammalian origin, e.g., human or murine, typically, of human origin. The nucleic acid molecule described herein is understood to refer to a modified form from the naturally-occurring sequence. In
20 some embodiments, the nucleic acid molecule is isolated or purified. In other embodiments, it is a recombinant molecule.

In one embodiment, the nucleic acid molecule has at least one, two, three introns, or up to all but one intron, deleted compared to the naturally-occurring sequence (e.g., the genomic sequence). For example, an intron that facilitates intron
25 read-through (IRT) can be deleted from the naturally-occurring sequence. In other embodiments, the nucleic acid molecule is modified by one or more of: re-arranging the intron/exon configuration (e.g., intron/exon 5' to 3' order); deleting a portion of

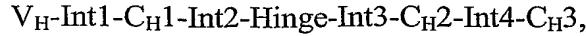
5 one or more introns; or replacing an intron or portion thereof with a heterologous
intron sequence, such that enhanced protein expression and/or reduction or
elimination of mis-spliced or intron read-through (IRT) by-product(s) occurs.

In a related embodiment, the nucleic acid molecule includes a nucleotide sequence (e.g., a human genomic sequence) encoding an antibody heavy chain or a fragment thereof. For example, the nucleotide sequence can include one or more nucleotide (e.g., exon) sequences encoding a heavy chain variable region, a hinge region, and a first, second, and third constant regions (e.g., C_H1, C_H2, C_H3) of an immunoglobulin subtype, e.g., an immunoglobulin G subtype (e.g., an IgG1, IgG2, IgG3, or IgG4 antibody subtype). Typically, the immunoglobulin subtype is from mammalian origin, e.g., murine or human. In one embodiment, a human IgG1 or IgG4, or a mutated version thereof is chosen. For example, the constant region of an immunoglobulin can be mutated to result in one or more of: increased stability, reduced effector function, or reduced complement fixation. In one embodiment, human IgG4 is mutated to increase stability, e.g., having a replacement at residue 241 from serine to proline to increase stability of the hinge region. In other embodiments, the constant region is mutated to reduce glycosylation.

In one embodiment, the nucleic acid molecule is modified to delete at least one intron that facilitates intron-read through of the sequence. For example, an intron between C_H2 and C_H3 of the immunoglobulin heavy chain constant region can be deleted. Examples of other heavy chain immunoglobulin introns that can be deleted individually or in combination include an intron between the heavy chain variable region and C_H1, an intron between C_H1 and the hinge region, and an intron between the hinge region and C_H2, of the immunoglobulin heavy chain constant region. Any combination of the preceding introns can be deleted, including a combination of two, three introns, or up to all but one intron, of the aforesaid introns. In some embodiments, three introns of the heavy chain constant region are deleted, for example, the intron between C_H1 and the hinge region, the intron between the hinge region and C_H2, and the intron between C_H2 and C_H3. The following exemplary combinations of intron deletions of a heavy chain immunoglobulin are also within the scope of the present invention: an intron between C_H1 and the hinge region, and an intron between C_H2 and C_H3; an intron between C_H1 and the hinge region, and an intron between the hinge region and C_H2; an intron between the hinge region and C_H2

- 5 and an intron between C_H2 and C_H3 of the immunoglobulin heavy chain constant region.

In some embodiments, the nucleic acid molecule includes a nucleotide sequence represented by the formula:



10 wherein V_H is a nucleotide sequence encoding a heavy chain variable region; C_H1, C_H2, and C_H3 are nucleotide sequences encoding the corresponding heavy chain constant region, e.g., a naturally-occurring or a mutated form of human IgG1 or IgG4 heavy chain gene;

15 Hinge is a nucleotide sequence encoding a hinge region of a heavy chain constant region, e.g., a naturally-occurring or a mutated form of human IgG1 or IgG4 heavy chain gene; and

Int1, Int2, Int3 and Int4 are introns from the heavy chain genomic sequence. In one embodiment, the intron between C_H2 and C_H3, represented herein as Int4 is deleted. In other embodiments, one, two, or typically three of the introns between 20 C_H1 and the hinge region, between the hinge region and C_H2, and/or between C_H2 and C_H3, represented herein as Int2, Int3 and Int4, are deleted. Additional schematic representations of the intron/exon arrangements of the heavy chain genomic sequence are shown in Figures 1, 5, and 7.

25 Typically, at least one intron is present in the nucleic acid molecule, for example, the intron between the heavy chain variable region and C_H1, represented herein as Int1. Examples of other heavy chain immunoglobulin introns that can be present individually or in combination include an intron between C_H1 and the hinge region; an intron between the hinge region and C_H2; and an intron between C_H2 and C_H3 of the immunoglobulin heavy chain constant region. It is often desirable to 30 include at least one intron in the modified nucleic acid molecule. Without being bound by theory, introns are believed to influence a number of events in the protein production process, including transcription rate, polyadenylation, mRNA export, translational efficiency, and mRNA decay.

In one embodiment, the nucleic acid molecule includes a nucleotide sequence 35 represented by the formula:



wherein V_H is a nucleotide sequence encoding a heavy chain variable region;

5 C_H1, C_H2, and C_H3 are nucleotide sequences encoding the corresponding heavy chain constant region, e.g., a naturally-occurring or mutated form of human IgG1 or IgG4 heavy chain gene;

10 Hinge is a nucleotide sequence encoding a hinge region of a heavy chain constant region, e.g., a naturally-occurring or mutated form of human IgG1 or IgG4 heavy chain gene; and

15 Int1, Int2 and Int3 are introns from the heavy chain genomic sequence. In one embodiment, the nucleotide sequence consists essentially of the constituents depicted above, e.g., without an intervening sequence that alters the structure or function.

20 In other embodiments, the nucleic acid molecule includes a nucleotide sequence represented by the formula:

$$V_H\text{-Int1-C}_H1\text{-Hinge-C}_H2\text{-C}_H3,$$

25 wherein V_H is a nucleotide sequence encoding a heavy chain variable region;

30 C_H1, C_H2, and C_H3 are nucleotide sequences encoding the corresponding heavy chain constant region, e.g., a naturally-occurring or mutated form of human IgG1 or IgG4 heavy chain gene;

35 Hinge is a nucleotide sequence encoding a hinge region of a heavy chain constant region, e.g., a naturally-occurring or mutated form of human IgG1 or IgG4 heavy chain gene; and

40 Int1 is an intron from the heavy chain genomic sequence. In one embodiment, the nucleotide sequence consists essentially of the constituents depicted above, e.g., without an intervening sequence that alters the structure or function.

45 The genomic nucleotide and corresponding amino acid sequences for human IgG1 are shown in Figure 8 (SEQ ID NO:1 and 2, respectively). Exons encoding C_H1, the hinge region, C_H2, and C_H3 are located at about nucleotides 231 to 524, 916 to 960, 1079 to 1408, and 1506 to 1829, of Figure 8 (SEQ ID NO:1), respectively. The Int1, Int2, Int3 and Int4 correspond to introns from the human IgG1 heavy chain genomic sequence located from about nucleotides 1 to 230, about nucleotides 525 to 915, about nucleotides 961 to 1078, and about nucleotides 1409 to 1505, of Figure 8 (SEQ ID NO:1), respectively.

50 The genomic nucleotide and corresponding amino acid sequences for mutated human IgG4 are shown in Figure 9 (SEQ ID NO:3 and 4 respectively). Exons encoding C_H1, the hinge region, C_H2, and C_H3 are located at about nucleotides 231 to

5 524, 916 to 952, 1071 to 1400, and 1498 to 1820, of Figure 9 (SEQ ID NO:3), respectively. Int1, Int2, Int3, and Int4 correspond to introns from the human IgG4 heavy chain genomic sequence located from about nucleotides 1 to 230, about nucleotides 525 to 916, about nucleotides 953 to 1070, and about nucleotides 1401 to 1497, of Figure 9 (SEQ ID NO:3), respectively.

10 Examples of modified nucleic acid molecules of the present invention include a human genomic heavy chain constant region sequence having a deletion of the intron between CH2 and CH3 of, human IgG1, corresponding to about nucleotides 1409 to 1505 of Figure 8 (SEQ ID NO:1), or of mutated human IgG4, corresponding to about nucleotides 1401 to 1497 of Figure 9 (SEQ ID NO:3). Examples of other
15 heavy chain immunoglobulin introns that can be deleted individually or in combination include an intron between the heavy chain variable region and CH1 of, human IgG1, corresponding to about nucleotides 1 to 230 of Figure 8 (SEQ ID NO:1), or mutated human IgG4, corresponding to about nucleotides 1 to 230 of Figure 9 (SEQ ID NO:3); an intron between CH1 and the hinge region of, human IgG1,
20 corresponding to about nucleotides 525 to 915 of Figure 8 (SEQ ID NO:1), or mutated human IgG4, corresponding to about nucleotides 525 to 916 of Figure 9 (SEQ ID NO:3); and an intron between the hinge region and CH2, of human IgG1, corresponding to about nucleotides 961 to 1078 of Figure 8 (SEQ ID NO:1), or mutated human IgG4, corresponding to about nucleotides 953 to 1070 of Figure 9
25 (SEQ ID NO:3). Any combination of the preceding introns can be deleted, including a combination of two, three, four introns, or up to all but one intron, of the aforesaid introns can be deleted. In some embodiments, three introns of the heavy chain constant region are deleted, for example, the intron between CH1 and the hinge region, between the hinge region and CH2, and between CH2 and CH3. In some
30 embodiments, the nucleic acid molecule includes one or more of the exonic nucleotide sequences, and one or more (but not all) of the intronic nucleotide sequences, for human IgG1 or IgG4 disclosed herein, or a sequence substantially identical thereto. In a related embodiment, the nucleic acid molecule has a deletion in one or more (but not all) of the intronic nucleotide sequences, for human IgG1 or
35 IgG4 disclosed herein, or a sequence substantially identical thereto.

In one embodiment, the modified nucleic acid molecule includes the nucleotide sequence encoding human IgG1 shown as Figure 10 (SEQ ID NO:5) or a

5 sequence substantially identical thereto (e.g., a sequence at least 85%, 90%, 95%, or 99% identical to SEQ ID NO:5, or having one, five, ten, fifty or more nucleotide changes compared to the nucleotide sequence of SEQ ID NO:5).

In another embodiment, the modified nucleic acid molecule includes the nucleotide sequence of modified human IgG4 shown as Figure 11 (SEQ ID NO:6) or
10 a sequence substantially identical thereto (e.g., a sequence at least 85%, 90%, 95%, or 99% identical to SEQ ID NO:6, or having one, five, ten, fifty or more nucleotide changes compared to the nucleotide sequence of SEQ ID NO:6).

The modified nucleic acid molecule can include a nucleotide sequences encoding a light and heavy chain antibody or immunoglobulin sequence. Such
15 sequences can be present in the same nucleic acid molecule (e.g., the same expression vector) or alternatively, can be expressed from separate nucleic acid molecules (e.g., separate expression vectors). Typically, the encoded antibody or immunoglobulins or fragments thereof can include at least one, and preferably two full-length heavy chains, and at least one, and preferably two light chains. Alternatively, the encoded
20 immunoglobulins or fragments thereof can include only an antigen-binding fragment (e.g., an Fab, F(ab')₂, Fv or a single chain Fv fragment). The antibody or fragment thereof can be a monoclonal or single specificity antibody. The antibody or fragment thereof can also be a human, humanized, chimeric, CDR-grafted, or *in vitro* generated antibody. In yet other embodiments, the antibody has a heavy chain constant region
25 chosen from, e.g., IgG1, IgG2, IgG3, IgG4, IgM, IgA1, IgA2, IgD, and IgE; more particularly, chosen from, e.g., IgG1, IgG2, IgG3, and IgG4. In another embodiment, the antibody has a light chain chosen from, e.g., kappa or lambda.

In another embodiment, the nucleic acid molecule includes a variable region, for example a humanized, chimeric, CDR-grafted, or *in vitro* generated variable
30 region. Typically, the variable region specifically binds to a predetermined antigen, e.g., an antigen associated with a disorder, e.g., a neurodegenerative or a malignant disorder.

In one embodiment, the disorder is a neurodegenerative disorder and the antibody binds to an amyloid protein, for example, an A β peptide (e.g., a human A β peptide). For example, the antibody can be a humanized antibody against an A β peptide having a heavy chain and light chain variable regions containing one or more complementarity determining regions (CDRs) from a murine antibody, e.g., the
35

5 mouse anti-A β 3D6 antibody. The variable region of the humanized antibody typically includes a human or substantially human framework region. In one embodiment, the nucleic acid molecule includes the heavy and light chain variable regions of the humanized anti-A β peptide antibody.

In another embodiment, the disorder is a malignant or cancerous disorder and
10 the antibody binds to a cell surface protein associated with malignant cells (e.g., a solid tumor cell) for example a 5T4 protein. 5T4 protein is a 72 kDa glycoprotein expressed widely in carcinomas, particularly colorectal and gastric metastatic cancers. In some embodiments, the antibody is a humanized antibody against a 5T4 protein having a heavy chain and light chain variable regions containing one or more
15 complementarity determining regions (CDRs) from a murine antibody. In one embodiment, the nucleic acid molecule includes the heavy and light chain variable regions of the humanized anti-5T4 antibody.

In another aspect, the invention features a vector (e.g., an expression vector) including one or more of the foregoing modified nucleic acid molecules. The vector
20 can additionally include a nucleotide sequence that enhances one or more of: replication, selection, mRNA transcription, mRNA stability, protein expression or protein secretion, in a host cell. For example, the vector may include nucleotide sequences responsible for replication or enhancer expression, enhancer promoter elements, nucleotide sequences encoding a leader sequence, a gene encoding a
25 selectable marker (e.g., DHFR), an internal ribosomal entry site sequence (IRES), and polyadenylation sequences).

In another aspect, the invention provides a cell, for example, a eukaryotic host cell, e.g., a mammalian host cell (e.g., a Chinese Hamster Ovary (CHO) cell), including one of the foregoing nucleic acid molecules and/or vectors, e.g., expression
30 vectors. The cell can be transiently or stably transfected with the nucleic acid sequences of the invention.

In another aspect, the invention provides a method for enhancing expression of recombinant proteins or peptides, e.g., antibodies, or expressing recombinant proteins or peptides, e.g., antibodies having reduced levels of (e.g., substantially free of) mis-spliced and/or intron read-through products, compared to a reference, e.g., a naturally occurring genomic sequence. The method includes introducing a nucleic acid
35 molecule as described herein into a host cell, e.g., a mammalian host cell (e.g., a CHO

5 cell); culturing said host cell under conditions that allow expression of the recombinant protein or peptide to produce a culture of host cells; and optionally, obtaining, e.g., purifying, the recombinant protein or peptide, from the culture of host cells (e.g., host cell supernatants).

The method can further include the steps of identifying (e.g., detecting and/or determining the level of) IRT or an IRT product, in a nucleic acid sample, e.g., an mRNA sample from the host cell, by contacting said sample with nucleic acid probes complementary to an intron and an adjacent exon sequence, or alternatively, complementary to adjacent exon sequences, under conditions that allow hybridization of the nucleic acid sample and the probes; detecting the resulting complex, e.g., by 10 PCR amplification of the probe sequences. Detection of a complex, e.g., a PCR amplified product, in the sample containing the nucleic acid probe complementary to the intron sequence is indicative of the occurrence IRT or the IRT product. The level of an IRT product can be quantified as described, e.g., in Example 1.

In another aspect, a method for producing an antibody or fragment thereof 20 having reduced (e.g., substantially devoid of) intron read-through (IRT) heavy chain by-product, compared to a standard reference, e.g., a naturally occurring genomic sequence, is provided. The method includes culturing a cell, e.g., a mammalian cell (e.g., a CHO cell) containing a nucleic acid molecule as described herein and, optionally, a nucleic acid encoding an antibody light chain, under conditions such that 25 the heavy and light chains are expressed and, optionally, operatively associate. The antibody or fragment thereof are, optionally, purified from the cell culture. Typically, the antibody, or fragment thereof, has reduced mis-spliced or intron read-through (IRT) heavy chain by-product.

The method can further include the steps of detecting and/or determining the 30 level of IRT, or an IRT product, in a sample, e.g., an mRNA sample from the host cell; contacting said sample with nucleic acid probes complementary to an intron and an adjacent exon sequence, or alternatively, complementary to adjacent exon sequences, under conditions that allow hybridization of the nucleic acid sample and the probes; detecting the resulting complex, e.g., by PCR amplification of the probe 35 sequences. Detection of a complex, e.g., a PCR amplified product, in the sample containing the nucleic acid probe complementary to the intron sequence is indicative

5 of the occurrence IRT, or the IRT product. The level of an IRT product can be quantified as described, e.g., in Example 1.

In another aspect, the invention provides a method of reducing intron read-through (IRT) antibody heavy chain by-product expressed from a genomic heavy chain sequence, by deleting at least one intron from said sequence, wherein said 10 intron facilitates IRT.

In another aspect, the invention features a method of identifying (e.g., detecting and/or determining the level of) IRT or an IRT product, in a sample, e.g., a nucleic acid sample. The method includes: obtaining a nucleic acid sample, e.g., an mRNA sample from a cell, e.g., a recombinant cell (e.g., a host cell as described 15 herein); contacting said nucleic acid sample with nucleic acid probes complementary to an intron and an adjacent exon sequence, or alternatively, complementary to adjacent exon sequences, under conditions that allow hybridization of the nucleic acid sample and the probes; detecting the resulting complex, e.g., by PCR amplification of the probe sequences. Detection of a complex, e.g., a PCR amplified product, in the 20 sample containing the nucleic acid probe complementary to the intron sequence is indicative of the occurrence IRT, or the IRT product. The level of an IRT product can be quantified as described, e.g., in Example 1.

In another aspect, the invention features an antibody (e.g., a recombinant antibody), or fragment thereof, having reduced (e.g., substantially free of) mis-spliced 25 and/or intron read-through products, compared to a reference, e.g., a naturally occurring genomic sequence, produced according to the methods disclosed herein. In one embodiment, the antibody or fragment thereof is a chimeric, humanized, CDR-grafted or an *in vitro* generated antibody. Typically, the antibody or fragment thereof has a variable region that specifically binds to a predetermined antigen, e.g., an 30 antigen associated with a disorder, e.g., a neurodegenerative or a malignant disorder.

In another aspect, the invention provides a composition, e.g., a pharmaceutical composition, containing recombinant proteins or peptides, e.g., antibodies, having reduced (e.g., substantially free of) mis-spliced and/or intron read-through products, compared to a reference, e.g., a naturally occurring genomic sequence, and a 35 pharmaceutically acceptable carrier. These compositions are suitable for therapeutic use, including, for example, treatment of neurodegenerative and malignant disorders.

5 Other features and advantages of the invention will be apparent from the following detailed description and claims.

Brief Description of the Drawings

10 FIG. 1 depicts the expected pre-mRNA transcribed from the expression vector containing the 3D6 IgG gene (top) as well as the correctly spliced mRNA (middle) and intron-read through mRNA (bottom).

15 FIG. 2 shows the nucleic acid sequence spanning the intron between the CH2 and CH3 constant regions (referred to as the fourth intron) of the 3D6 heavy-chain expression vector indicating genomic 5' and 3' splice junctions (SEQ ID NO:7). Also shown is the predicted partial amino acid sequence of the polypeptides derived from correctly (SEQ ID NO:8) and incorrectly (SEQ ID NO:9) spliced mRNA. The RNA splice junctions are indicated by a solid double line.

20 FIG. 3 is a schematic representation of the quantitative-polymerase chain reaction (Q-PCR) probes used to evaluate total levels of 3D6 heavy chain gene transcription (levels of CH2 containing mRNA transcript) and levels of intron 4 read-through transcription.

FIG. 4 is a bar graph demonstrating the increased accumulation of intron 4 containing transcripts in response to time in culture and protein expression induction.

25 FIG. 5 provides drawings of the genomic arrangement of 3D6 introns and exons and the modified arrangement used in an expression vector developed to resolve intron read through transcription.

FIG. 6 shows reverse-phase high-performance liquid chromatography (RP-HPLC) chromatograms demonstrating the lack of intron read through heavy chain by-products in a cell line transformed with modified expression vectors.

30 FIG. 7 depicts the arrangement of introns and exons in a heavy chain genomic construct, a construct, the construct with the last three intronic sequences deleted, and the cDNA construct containing no introns.

FIG. 8 shows the genomic nucleotide and corresponding amino acid sequences for human IgG1 are shown in (SEQ ID NO:1 and 2, respectively). Exons encoding 35 C_H1, the hinge region, C_H2, and C_H3 are located at about nucleotides 231 to 524, 916 to 960, 1079 to 1408, and 1506 to 1829, respectively (SEQ ID NO:1). The Int1, Int2, Int3 and Int4 correspond to introns from the human IgG1 heavy chain genomic

5 sequence located from about nucleotides 1 to 230, about nucleotides 525 to 915, about nucleotides 961 to 1078, and about nucleotides 1409 to 1505, respectively (SEQ ID NO:1).

FIG. 9 shows the genomic nucleotide and corresponding amino acid sequences for human IgG4 are shown in (SEQ ID NO:3 and 4, respectively). Exons encoding 10 C_H1 , the hinge region, C_H2 , and C_H3 are located at about nucleotides 231 to 524, 916 to 952, 1071 to 1400, and 1498 to 1820, respectively, (SEQ ID NO:3). Int1, Int2, Int3, and Int4 correspond to introns from the human IgG4 heavy chain genomic sequence located from about nucleotides 1 to 230, about nucleotides 525 to 916, about nucleotides 953 to 1070, and about nucleotides 1401 to 1497, respectively (SEQ ID 15 NO:3).

FIG. 10 shows the genomic nucleotide sequence of human IgG1 (SEQ ID NO:5) having the intron between CH2 and CH3 of the constant region deleted.

FIG. 11 shows the genomic nucleotide sequence of modified human IgG4 (SEQ ID NO:6) having the following intron deletions: intron between C_H1 and hinge, intron between hinge and C_H2 , and intron between C_H2 and C_H3 . 20

Detailed Description of the Invention

A number of approaches may be taken in the design and construction of expression vectors, and the process typically requires substantial trial and error 25 experimentation before reasonable levels of a protein are produced. A significant consideration in the design process concerns the use of intron sequences in the construction of the vector. In one approach, an entire gene sequence may be utilized as it occurs naturally – containing the full complement of both intronic and exonic sequences. In such a case, it is expected that post-transcriptional splicing machinery 30 within the cell will excise intronic sequences to yield a mature mRNA containing only exonic sequences of the gene. A second approach is to utilize sequence corresponding to the cDNA of the gene only. In this case, it is predicted that no 35 splicing events occur and the pre-mRNA sequence is substantially the same as the mRNA sequence in protein coding content. In yet a third case, vector construction involves the selection and placement of introns not normally associated with the original gene sequence.

5 The effect of intronic sequences on the expression of genes within the context
of a vector is incompletely understood. It has been reported that introns may effect a
number of events in the process of protein production including transcription rate,
polyadenylation, mRNA export, translational efficiency, and mRNA decay (Nott *et al*
(2003) *RNA* 9:607-617). Within the context of mRNA expression, there has been no
10 bright line of predictability regarding the result of an intron on the yield of protein
from a vector. For example, it has been variously reported that including various
intronic sequences can cause large increases in expression, have no effect, or reduce
mRNA expression (Berg *et al.* (1988) *Mol. Cell. Biol.* 8:4395-4405; Bourdon *et al.*
(2001) *EMBO Rep.* 2:394-398). Since most higher eukaryotic genes contain introns,
15 the development of a system which may be used to predictably express intron-
containing genes at high levels and with close fidelity to the exonic sequences of the
gene in the absence of unwanted read-through by products is obviously an aid to the
predictable development of protein expression systems.

While the unpredictability associated with intronic sequences poses a hurdle to
20 reliable expression vector design, a significant design benefit can be realized when the
protein of interest has a modular form which is amenable to genetic engineering
techniques. Antibodies provide one such example wherein the inclusion of intronic
sequences facilitates expression vector design.

Certain terms used in the specification and claims are defined below.

25 The phrase “intron read-through” (“IRT”) denotes the process whereby
aberrant splicing of a pre-mRNA transcript yields a protein or peptide of alternate size
or amino acid constituency. Varying results may occur concerning the ultimate
protein produced from the mis-spliced transcript. For example, a larger than predicted
protein or a protein with an incorrect stop codon may occur, in which case the protein
30 may be longer or shorter than predicted, respectively. Further, the protein may also
have incorrect or additional residues facilitating protein modification for
glycosylation, myristylation, phosphorylation, ubiquitination, or other post-
translational modifications.

The term “intron read-through by-product” refers to proteins or peptides that
35 are translated from aberrantly-spliced mRNA resulting from intron read-through, e.g.,
proteins of unpredicted size or amino acid constituency. Intron read-through by
products may be shorter or longer than the polypeptides predicted by the genes known

5 amino acid sequence and/or predicted by the cDNA of the gene. Intron read-through by products may also have apparent molecular weights differing from the accepted molecular weight of proteins arising from the correctly spliced mRNA of the gene. Further, the term “intron read-through by-products” includes proteins that occur from proteolytic processing events not normally associated with the protein of interest, said
10 proteolytic processing arising potentially from frame shifted protein products due to read through of an intron-exon-junction.

The term “heavy chain by-product” refers to polypeptides that are translated from aberrantly spliced immunoglobulin heavy chain mRNA resulting from intron read-through, e.g. a heavy chain protein of unpredicted size or amino acid
15 constituency. Heavy chain byproducts may be shorter or longer than the polypeptide predicted by the immunoglobulin gene’s known amino acid sequence and/or predicted by the cDNA of the gene. Heavy chain by-products may also have apparent molecular weights differing from the accepted molecular weight of proteins arising from the correctly spliced mRNA of the heavy chain gene. Further, the term “heavy
20 chain by-products” includes polypeptides that occur from proteolytic processing events not normally associated with the protein.

The phrase “naturally-occurring sequence” or “naturally-occurring genomic sequence” refers to the intronic and exonic organization of a gene found in its natural or native state. The naturally-occurring sequence can be found in, e.g., its natural
25 chromosomal location or cloned into a vector, so long as the intronic and exonic organization of the sequence is retained.

The term “immunoglobulin” or “antibody” (used interchangeably herein) refers to a protein having a four-polypeptide chain structure consisting of two heavy and two light chains, said chains being stabilized, for example, by interchain disulfide bonds, wherein the immunoglobulin or antibody has the ability to specifically bind an
30 antigen.

The term “single-chain immunoglobulin” or “single-chain antibody” (used interchangeably herein) refers to a protein having a two-polypeptide chain structure consisting of a heavy and a light chain, said chains being stabilized, for example, by
35 interchain peptide linkers, wherein the immunoglobulin or antibody has the ability to specifically bind antigen.

5 The term “immunoglobulin or antibody domain” refers to a globular region
within a heavy or light chain polypeptide including peptide loops (*e.g.*, including 3 to
4 peptide loops) stabilized, for example, by β -pleated sheet and/or intrachain disulfide
bond. Domains are further referred to herein as “constant” or “variable” wherein the
term “constant” refers to the relative lack of sequence variation within the domains of
10 various class members in the case of a “constant” domain and wherein the term
“variable” refers to the significant variation within the domains of various class
members in the case of a “variable” domain. Antibody or polypeptide “domains” are
often referred to interchangeably in the art as antibody or polypeptide “regions.” The
“constant” domains of an antibody light chain are referred to interchangeably as “light
15 chain constant regions,” “light chain constant domains,” “CL” regions or “CL”
domains. The “constant” domains of an antibody heavy chain are referred to
interchangeably as “heavy chain constant regions,” “heavy chain constant domains,”
“CH” regions or “CH” domains. The “variable” domains of an antibody light chain
are referred to interchangeably as “light chain variable regions,” “light chain variable
20 domains,” “VL” regions or “VL” domains. The “variable” domains of an antibody
heavy chain are referred to interchangeably as “heavy chain constant regions,” “heavy
chain constant domains,” “VH” regions or “VH” domains.

The term “region” can also refer to a part or portion of an antibody chain or
antibody chain domain (*e.g.*, a part or portion of a heavy or light chain or a part or
25 portion of a constant or variable domain, as defined herein), as well as more discrete
parts or portions of said chains or domains. For example, light and heavy chains or
light and heavy chain variable domains include “complementarity determining
regions” or “CDRs” interspersed among “framework regions” or “FRs”, as defined
herein.

30 Immunoglobulins or antibodies can exist in monomeric or polymeric form, for
example, IgM antibodies, which exist in pentameric form, and/or IgA antibodies,
which exist in monomeric, dimeric or multimeric form. The term “fragment” refers to
a part or portion of an antibody or antibody chain including fewer amino acid residues
35 than an intact or complete antibody or antibody chain. Fragments can be obtained *via*
chemical or enzymatic treatment of an intact or complete antibody or antibody chain.
Fragments can also be obtained by recombinant means. Exemplary fragments include
Fab, Fab’, F(ab’)₂, Fabc, and/or Fv fragments. The term “antigen-binding fragment”

5 refers to a polypeptide fragment of an immunoglobulin or antibody that binds antigen or competes with intact antibody (*i.e.*, with the intact antibody from which they were derived) for antigen binding (*i.e.*, specific binding).

The term “conformation” refers to the tertiary structure of a protein or polypeptide (*e.g.*, an antibody, antibody chain, domain or region thereof). For 10 example, the phrase “light (or heavy) chain conformation” refers to the tertiary structure of a light (or heavy) chain variable region, and the phrase “antibody conformation” or “antibody fragment conformation” refers to the tertiary structure of an antibody or fragment thereof. The term “conformation” may also refer to quaternary structures resulting from the three dimensional relationship of one or 15 several proteins or peptide chains. In relation to antigenic determinants, the phrase “conformational epitope” refers to an antigenic determinant including a specific spatial arrangement of amino acids within one or several proteins existing in close apposition. Considering the multifunctional nature of antibodies (*i.e.* the ability of IgG molecules to bind several epitopes concomitantly on more than one protein 20 molecule), antibodies can be considered as having the innate ability to bind conformational epitopes comprised by several amino acid chains. For example, the deposition of A β to form plaques provides a conformational epitope in which one antibody may bind several closely positioned A β peptides.

Binding fragments are produced by recombinant DNA techniques, or by 25 enzymatic or chemical cleavage of intact immunoglobulins. Binding fragments include Fab, Fab', F(ab')₂, Fabc, Fv, single chains, and single-chain antibodies. Other than “bispecific” or “bifunctional” immunoglobulins or antibodies, an immunoglobulin or antibody is understood to have each of its binding sites identical. A “bispecific” or “bifunctional antibody” is an artificial hybrid antibody having two 30 different heavy/light chain pairs and two different binding sites. Bispecific antibodies can be produced by a variety of methods including fusion of hybridomas or linking of Fab' fragments. See, *e.g.*, Songsivilai & Lachmann, *Clin. Exp. Immunol.* 79:315-321 (1990); Kostelny *et al.*, *J. Immunol.* 148, 1547-1553 (1992).

The term “significant identity” means that two sequences, *e.g.*, two 35 polypeptide sequences, when optimally aligned, such as by the programs GAP or BESTFIT using default gap weights, share at least 50-60% sequence identity, preferably at least 60-70% sequence identity, more preferably at least 70-80%

5 sequence identity, more preferably at least 80-90% identity, even more preferably at
least 90-95% identity, and even more preferably at least 95% sequence identity or
more (e.g., 99% sequence identity or more). The term “substantial identity” or
“substantially identical” means that two sequences, e.g., two polypeptide sequences,
when optimally aligned, such as by the programs GAP or BESTFIT using default gap
10 weights, share at least 80-90% sequence identity, preferably at least 90-95% sequence
identity, and more preferably at least 95% sequence identity or more (e.g., 99%
sequence identity or more). For sequence comparison, typically one sequence acts as
a reference sequence, to which test sequences are compared. When using a sequence
comparison algorithm, test and reference sequences are input into a computer,
15 subsequence coordinates are designated, if necessary, and sequence algorithm
program parameters are designated. The sequence comparison algorithm then
calculates the percent sequence identity for the test sequence(s) relative to the
reference sequence, based on the designated program parameters.

Optimal alignment of sequences for comparison can be conducted, e.g., by the
20 local homology algorithm of Smith & Waterman, *Adv. Appl. Math.* 2:482 (1981), by
the homology alignment algorithm of Needleman & Wunsch, *J. Mol. Biol.* 48:443
(1970), by the search for similarity method of Pearson & Lipman, *Proc. Nat'l. Acad.
Sci. USA* 85:2444 (1988), by computerized implementations of these algorithms
(GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software
25 Package, Genetics Computer Group, 575 Science Dr., Madison, WI), or by visual
inspection (see generally Ausubel et al., *Current Protocols in Molecular Biology*).
One example of algorithm that is suitable for determining percent sequence identity
and sequence similarity is the BLAST algorithm, which is described in Altschul et al.,
J. Mol. Biol. 215:403 (1990). Software for performing BLAST analyses is publicly
30 available through the National Center for Biotechnology Information (publicly
accessible through the National Institutes of Health NCBI internet server). Typically,
default program parameters can be used to perform the sequence comparison,
although customized parameters can also be used. For amino acid sequences, the
BLASTP program uses as defaults a wordlength (W) of 3, an expectation (E) of 10,
35 and the BLOSUM62 scoring matrix (see Henikoff & Henikoff, *Proc. Natl. Acad. Sci.
USA* 89:10915 (1989)).

5 Preferably, residue positions which are not identical differ by conservative amino acid substitutions. For purposes of classifying amino acids substitutions as conservative or nonconservative, amino acids are grouped as follows: Group I (hydrophobic sidechains): leu, met, ala, val, leu, ile; Group II (neutral hydrophilic side chains): cys, ser, thr; Group III (acidic side chains): asp, glu; Group IV (basic side chains): asn, gln, his, lys, arg; Group V (residues influencing chain orientation): gly, pro; and Group VI (aromatic side chains): trp, tyr, phe. Conservative substitutions involve substitutions between amino acids in the same class. Non-conservative substitutions constitute exchanging a member of one of these classes for a member of another.

15

Antibodies

The methodologies of the present invention are applicable in a variety of antibody production processes where unwanted or undesirable by-products are detected. In particular, the methodologies are applicable in production of 20 recombinant antibodies, such as chimeric and humanized monoclonal antibodies, where the sequence of the antibody being produced is known.

The term “humanized immunoglobulin” or “humanized antibody” refers to an immunoglobulin or antibody that includes at least one humanized immunoglobulin or antibody chain (*i.e.*, at least one humanized light or heavy chain). The term 25 “humanized immunoglobulin chain” or “humanized antibody chain” (*i.e.*, a “humanized immunoglobulin light chain” or “humanized immunoglobulin heavy chain”) refers to an immunoglobulin or antibody chain (*i.e.*, a light or heavy chain, respectively) having a variable region that includes a variable framework region substantially from a human immunoglobulin or antibody and complementarity 30 determining regions (CDRs) (*e.g.*, at least one CDR, preferably two CDRs, more preferably three CDRs) substantially from a non-human immunoglobulin or antibody, and further includes constant regions (*e.g.*, at least one constant region or portion thereof, in the case of a light chain, and preferably three constant regions in the case of a heavy chain). The term “humanized variable region” (*e.g.*, “humanized light 35 chain variable region” or “humanized heavy chain variable region”) refers to a variable region that includes a variable framework region substantially from a human

- 5 immunoglobulin or antibody and complementarity determining regions (CDRs) substantially from a non-human immunoglobulin or antibody.

The phrase "substantially from a human immunoglobulin or antibody" or "substantially human" means that, when aligned to a human immunoglobulin or antibody amino sequence for comparison purposes, the region shares at least 80-90%,
10 90-95%, or 95-99% identity (*i.e.*, local sequence identity) with the human framework or constant region sequence, allowing, for example, for conservative substitutions, consensus sequence substitutions, germline substitutions, back-mutations, and the like. The introduction of conservative substitutions, consensus sequence substitutions, germline substitutions, back-mutations, and the like, is often referred to
15 as "optimization" of a humanized antibody or chain. The phrase "substantially from a non-human immunoglobulin or antibody" or "substantially non-human" means having an immunoglobulin or antibody sequence at least 80-95%, preferably at least 90-95%, more preferably, 96%, 97%, 98%, or 99% identical to that of a non-human organism, *e.g.*, a non-human mammal.

20 Accordingly, all regions or residues of a humanized immunoglobulin or antibody, or of a humanized immunoglobulin or antibody chain, except possibly the CDRs, are substantially identical to the corresponding regions or residues of one or more native human immunoglobulin sequences. The term "corresponding region" or "corresponding residue" refers to a region or residue on a second amino acid or nucleotide sequence which occupies the same (*i.e.*, equivalent) position as a region or residue on a first amino acid or nucleotide sequence, when the first and second sequences are optimally aligned for comparison purposes.

25 Preferably, humanized immunoglobulins or antibodies bind antigen with an affinity that is within a factor of three, four, or five of that of the corresponding non-humanized antibody. For example, if the non-humanized antibody has a binding affinity of 10^9 M^{-1} , humanized antibodies will have a binding affinity of at least $3 \times 10^9 \text{ M}^{-1}$, $4 \times 10^9 \text{ M}^{-1}$, or $5 \times 10^9 \text{ M}^{-1}$. When describing the binding properties of an immunoglobulin or antibody chain, the chain can be described based on its ability to "direct antigen (*e.g.*, A β or 5T4) binding." A chain is said to "direct antigen binding" when it confers upon an intact immunoglobulin or antibody (or antigen binding fragment thereof) a specific binding property or binding affinity. A mutation (*e.g.*, a back-mutation) is said to substantially affect the ability of a heavy or light chain to

- 5 direct antigen binding if it affects (*e.g.*, decreases) the binding affinity of an intact immunoglobulin or antibody (or antigen binding fragment thereof) comprising said chain by at least an order of magnitude compared to that of the antibody (or antigen binding fragment thereof) comprising an equivalent chain lacking said mutation. A mutation “does not substantially affect (*e.g.*, decrease) the ability of a chain to direct
10 antigen binding” if it affects (*e.g.*, decreases) the binding affinity of an intact immunoglobulin or antibody (or antigen binding fragment thereof) comprising said chain by only a factor of two, three, or four of that of the antibody (or antigen binding fragment thereof) comprising an equivalent chain lacking said mutation.

The term “chimeric immunoglobulin” or antibody refers to an
15 immunoglobulin or antibody whose variable regions derive from a first species and whose constant regions derive from a second species. Chimeric immunoglobulins or antibodies can be constructed, for example by genetic engineering, from immunoglobulin gene segments belonging to different species. The terms “humanized immunoglobulin” or “humanized antibody” are not intended to
20 encompass chimeric immunoglobulins or antibodies, as defined herein. Although humanized immunoglobulins or antibodies are chimeric in their construction (*i.e.*, comprise regions from more than one species of protein), they include additional features (*i.e.*, variable regions comprising donor CDR residues and acceptor framework residues) not found in chimeric immunoglobulins or antibodies, as defined
25 herein.

Such chimeric and humanized monoclonal antibodies can be produced by recombinant DNA techniques known in the art, for example using methods described in Robinson *et al.* International Application No. PCT/US86/02269; Akira, *et al.* European Patent Application 184,187; Taniguchi, M., European Patent Application 30 171,496; Morrison *et al.* European Patent Application 173,494; Neuberger *et al.* PCT International Publication No. WO 86/01533; Cabilly *et al.* U.S. Patent No. 4,816,567; Cabilly *et al.* European Patent Application 125,023; Better *et al.* (1988) *Science* 240:1041-1043; Liu *et al.* (1987) *Proc. Natl. Acad. Sci. USA* 84:3439-3443; Liu *et al.* (1987) *J. Immunol.* 139:3521-3526; Sun *et al.* (1987) *Proc. Natl. Acad. Sci. USA* 84:214-218; Nishimura *et al.* (1987) *Canc. Res.* 47:999-1005; Wood *et al.* (1985) *Nature* 314:446-449; and Shaw *et al.* (1988) *J. Natl. Cancer Inst.* 80:1553-1559);
35 Morrison, S. L. (1985) *Science* 229:1202-1207; Oi *et al.* (1986) *BioTechniques*

5 4:214; Winter U.S. Patent 5,225,539; Jones *et al.* (1986) *Nature* 321:552-525;
Verhoeven *et al.* (1988) *Science* 239:1534; and Beidler *et al.* (1988) *J. Immunol.*
141:4053-4060.

Monoclonal, chimeric and humanized antibodies, which have been modified,
e.g., by deleting, adding, or substituting other portions of the antibody, e.g., the
10 constant region, are also within the scope of the invention. For example, an antibody
can be modified as follows: (i) by replacing the constant region with another constant
region, e.g., a constant region meant to increase half-life, stability or affinity of the
antibody, or a constant region from another species or antibody class; or (ii) by
modifying one or more amino acids in the constant region to alter, for example, the
15 number of glycosylation sites, effector cell function, Fc receptor (FcR) binding,
complement fixation, among others. Methods for altering an antibody constant region
are known in the art. Antibodies with altered function, e.g. altered affinity for an
effector ligand, such as FcR on a cell, or the C1 component of complement can be
produced by replacing at least one amino acid residue in the constant portion of the
20 antibody with a different residue (see e.g., EP 388,151 A1, U.S. Pat. No. 5,624,821
and U.S. Pat. No. 5,648,260, the contents of all of which are hereby incorporated by
reference). Similar type of alterations could be described which if applied to the
murine, or other species immunoglobulin would reduce or eliminate these functions.

For example, it is possible to alter the affinity of an Fc region of an antibody
25 (e.g., an IgG, such as a human IgG) for an FcR (e.g., Fc gamma R1), or for C1q
binding by replacing the specified residue(s) with a residue(s) having an appropriate
functionality on its side chain, or by introducing a charged functional group, such as
glutamate or aspartate, or perhaps an aromatic non-polar residue such as
phenylalanine, tyrosine, tryptophan or alanine (see e.g., U.S. Pat. No. 5,624,821).

30

Human Antibodies from Transgenic Animals and Phage Display

Alternatively, it is now possible to produce transgenic animals (e.g., mice) that
are capable, upon immunization, of producing a full repertoire of human antibodies in
the absence of endogenous immunoglobulin production. For example, it has been
35 described that the homozygous deletion of the antibody heavy-chain joining region
(J_H) gene in chimeric and germ-line mutant mice results in complete inhibition of
endogenous antibody production. Transfer of the human germ-line immunoglobulin

5 gene array in such germ-line mutant mice results in the production of human antibodies upon antigen challenge. See, e.g., U.S. Patent Nos. 6,150,584; 6,114,598; and 5,770,429.

Fully human antibodies can also be derived from phage-display libraries (Hoogenboom *et al.*, J. Mol. Biol., 227:381 (1991); Marks *et al.*, J. Mol. Biol., 10 222:581-597 (1991)).

Bispecific Antibodies, Antibody Fusion Polypeptides, and Single-Chain Antibodies

Bispecific antibodies (BsAbs) are antibodies that have binding specificities for at least two different epitopes. Such antibodies can be derived from full length 15 antibodies or antibody fragments (e.g. F(ab)'₂ bispecific antibodies). Methods for making bispecific antibodies are known in the art. Traditional production of full length bispecific antibodies is based on the coexpression of two immunoglobulin heavy chain-light chain pairs, where the two chains have different specificities (Millstein *et al.*, Nature, 305:537-539 (1983)). Because of the random assortment of 20 immunoglobulin heavy and light chains, these hybridomas (quadromas) produce a potential mixture of different antibody molecules (see, WO 93/08829 and in Traunecker *et al.*, EMBO J., 10:3655-3659 (1991)).

Bispecific antibodies also include cross-linked or "heteroconjugate" antibodies. For example, one of the antibodies in the heteroconjugate can be coupled 25 to avidin, the other to biotin or other payload. Heteroconjugate antibodies may be made using any convenient cross-linking methods. Suitable cross-linking agents are well known in the art, and are disclosed in U.S. Pat. No. 4,676,980, along with a number of cross-linking techniques.

In yet another embodiment, the antibody can be fused, chemically or 30 genetically, to a payload domain, for example, an immunotoxin to produce an antibody fusion polypeptide. Such payloads include, for example, immunotoxins, chemotherapeutics, and radioisotopes, all of which are well-known in the art.

Single chain antibodies are also suitable for stabilization according to the invention. The fragments comprise a heavy-chain variable domain (VH) connected to 35 a light-chain variable domain (VL) with a linker, which allows each variable region to interface with each other and recreate the antigen binding pocket of the parent

5 antibody from which the VL and VH regions are derived. See Gruber *et al.*, J.
10 Immunol., 152:5368 (1994).

Anti-A β Antibodies

Generally, the antibodies of the present invention include antibodies for
10 treating amyloidogenic diseases, in particular, Alzheimer's Disease, by targeting A β
peptide.

The term "amyloidogenic disease" includes any disease associated with (or
caused by) the formation or deposition of insoluble amyloid fibrils. Exemplary
amyloidogenic diseases include, but are not limited to, systemic amyloidosis,
15 Alzheimer's disease, mature onset diabetes, Parkinson's disease, Huntington's
disease, fronto-temporal dementia, and the prion-related transmissible spongiform
encephalopathies (kuru and Creutzfeldt-Jacob disease in humans and scrapie and BSE
in sheep and cattle, respectively). Different amyloidogenic diseases are defined or
characterized by the nature of the polypeptide component of the fibrils deposited. For
20 example, in subjects or patients having Alzheimer's disease, β -amyloid protein (*e.g.*,
wild-type, variant, or truncated β -amyloid protein) is the characterizing polypeptide
component of the amyloid deposit. Accordingly, Alzheimer's disease is an example
of a "disease characterized by deposits of A β " or a "disease associated with deposits
of A β ," *e.g.*, in the brain of a subject or patient. The terms " β -amyloid protein," " β -
25 amyloid peptide," " β -amyloid," "A β ," and "A β peptide" are used interchangeably
herein.

An "immunogenic agent" or "immunogen" is capable of inducing an immunological
response against itself on administration to a mammal, optionally in conjunction with
an adjuvant.

30 The terms "A β antibody," "anti A β antibody," and "anti A β " are used
interchangeably herein to refer to an antibody that binds to one or more epitopes or
antigenic determinants of APP, A β protein, or both. Exemplary epitopes or antigenic
determinants can be found within the human amyloid precursor protein (APP), but are
preferably found within the A β peptide of APP. Multiple isoforms of APP exist, for
35 example APP⁶⁹⁵, APP⁷⁵¹, and APP⁷⁷⁰. Amino acids within APP are assigned numbers
according to the sequence of the APP⁷⁷⁰ isoform (see *e.g.*, GenBank Accession No.
P05067). A β (also referred to herein as beta amyloid peptide and A beta) peptide is a

- 5 ~4-kDa internal fragment of 39-43 amino acids of APP (A β 39, A β 40, A β 41, A β 42, and A β 43). A β 40, for example, consists of residues 672-711 of APP and A β 42 consists of residues 672-713 of APP. As a result of proteolytic processing of APP by different secretase enzymes *iv vivo* or *in situ*, A β is found in both a “short form,” 40 amino acids in length, and a “long form,” ranging from 42-43 amino acids in length.
- 10 Epitopes or antigenic determinants can be located within the N-terminus of the A β peptide and include residues within amino acids 1-10 of A β , preferably from residues 1-3, 1-4, 1-5, 1-6, 1-7, 2-7, 3-6, or 3-7 of A β 42 or within residues 2-4, 5, 6, 7, or 8 of A β , residues 3-5, 6, 7, 8, or 9 of A β , or residues 4-7, 8, 9, or 10 of A β 42. “Central” epitopes or antigenic determinants are located within the central or mid-portion of the
- 15 A β peptide and include residues within amino acids 16-24, 16-23, 16-22, 16-21, 19-21, 19-22, 19-23, or 19-24 of A β . “C-terminal” epitopes or antigenic determinants are located within the C-terminus of the A β peptide and include residues within amino acids 33-40, 33-41, or 33-42 of A β .

In various embodiments, an A β antibody is end-specific. As used herein, the term “end-specific” refers to an antibody which specifically binds to the N-terminal or C-terminal residues of an A β peptide but that does not recognize the same residues when present in a longer A β species comprising the residues or in APP.

In various embodiments, an A β antibody is “C-terminus-specific.” As used herein, the term “C terminus-specific” means that the antibody specifically recognizes a free C-terminus of an A β peptide. Examples of C terminus-specific A β antibodies include those that: recognize an A β peptide ending at residue 40, but do not recognize an A β peptide ending at residue 41, 42, and/or 43; recognize an A β peptide ending at residue 42, but do not recognize an A β peptide ending at residue 40, 41, and/or 43; etc.

- 30 In one embodiment, the antibody may be a 3D6 antibody or variant thereof, or a 10D5 antibody or variant thereof, both of which are described in U.S. Patent Publication No. 2003/0165496A1, U.S. Patent Publication No. 2004/0087777A1, International Patent Publication No. WO02/46237A3. Description of 3D6 and 10D5 can also be found, for example, in International Patent Publication No.
- 35 WO02/088306A2 and International Patent Publication No. WO02/088307A2. 3D6 is a monoclonal antibody (mAb) that specifically binds to an N-terminal epitope located

5 in the human β -amyloid peptide, specifically, residues 1-5. By comparison, 10D5 is a mAb that specifically binds to an N-terminal epitope located in the human β -amyloid peptide, specifically, residues 3-6. In another embodiment, the antibody may be a 12B4 antibody or variant thereof, as described in U.S. Patent Publication No. 20040082762A1 and International Patent Publication No. WO03/077858A2. 12B4 is
10 a mAb that specifically binds to an N-terminal epitope located in the human β -amyloid peptide, specifically, residues 3-7. In yet another embodiment, the antibody may be a 12A11 antibody or a variant thereof, as described in U.S. Patent Application No. 10/858,855 and International Patent Application No. PCT/US04/17514. 12A11 is a mAb that specifically binds to an N-terminal epitope located in the human β -
15 amyloid peptide, specifically, residues 3-7. In yet another embodiment, the antibody may be a 266 antibody as described in U.S. Patent Application No. 10/789,273, and International Patent Application No. WO01/62801A2. Antibodies designed to specifically bind to C-terminal epitopes located in human β -amyloid peptide, for use in the present invention include, but are not limited to, 369.2B, as described in U.S.
20 Patent No. 5,786,160.

In exemplary embodiments, the antibody is a humanized anti A β peptide 3D6 antibody that selectively binds A β peptide. More specifically, the humanized anti A β peptide 3D6 antibody is designed to specifically bind to an NH₂-terminal epitope located in the human β -amyloid 1-40 or 1-42 peptide found in plaque deposits in the
25 brain (e.g., in patients suffering from Alzheimer's disease).

Anti-5T4 Antibodies

The 5T4 antigen has been previously characterized (see e.g., WO 89/07947). The full nucleic acid sequence of human 5T4 is known (Myers *et al.* (1994) *J Biol
30 Chem* 169: 9319-24 and GenBank at Accession No. Z29083). The sequence for 5T4 antigen from other species is also known, for example, murine 5T4 (WO00/29428), canine 5T4 (WO01/36486) or feline 5T4 (US 05/0100958).

Human 5T4 is a glycoprotein of about 72 kDa expressed widely in carcinomas, but having a highly restricted expression pattern in normal adult tissues.
35 It appears to be strongly correlated to metastasis in colorectal and gastric cancer. Expression of the 5T4 antigen is also found at high frequency in breast and ovarian cancers (Starzynska *et al.* (1998) *Eur. J. Gastroenterol. Hepatol.* 10:479-84;

- 5 Starzynska *et al.* (1994) *Br. J. Cancer* 69:899-902; Starzynska et al. (1992) *Br. J. Cancer* 66:867-9). 5T4 has been proposed as a marker, with possible mechanistic involvement, for tumor progression and metastasis potential (Carsberg *et al.* (1996) *Int J Cancer* 68:84-92). 5T4 has also been proposed for use as an immunotherapeutic agent (see WO 00/29428). Antigenic peptides of 5T4 are disclosed in, e.g., US
10 05/0100958, the contents of which are incorporated by reference.

Several pending applications relate generally to nucleic acids encoding the anti-5T4 monoclonal antibody, vectors and host cells thereof, for example, U.S. Application Publication Nos. 2003/0018004 and 2005/0032216 and U.S. Application Serial No. 10/016,686. A provisional patent application pertaining generally to the
15 humanized anti-5T4 H8 monoclonal antibodies and calicheamicin conjugates thereof, as well as methods of treatment using these calicheamicin conjugates has been filed (U.S. Application Serial No. 60/608,494). The contents of all of these applications are incorporated by reference herein in their entirety.

20 Fc Fusions

In some embodiments, the nucleic acid molecules of the invention encode a fusion or a chimeric protein. The fusion protein can include a targeting moiety, e.g., a soluble receptor fragment or a ligand, and an immunoglobulin chain, an Fc fragment, a heavy chain constant regions of the various isotypes, including: IgG1, IgG2, IgG3,
25 IgG4, IgM, IgA1, IgA2, IgD, and IgE). For example, the fusion protein can include the extracellular domain of a receptor, and, e.g., fused to, a human immunoglobulin Fc chain (e.g., human IgG, e.g., human IgG1 or human IgG4, or a mutated form thereof). In one embodiment, the human Fc sequence has been mutated at one or more amino acids, e.g., mutated at residues 254 and 257 from the wild type sequence
30 to reduce Fc receptor binding. The fusion proteins may additionally include a linker sequence joining the first moiety to the second moiety, e.g., the immunoglobulin fragment. For example, the fusion protein can include a peptide linker, e.g., a peptide linker of about 4 to 20, more preferably, 5 to 10, amino acids in length; the peptide linker is 8 amino acids in length. For example, the fusion protein can include a
35 peptide linker having the formula (Ser-Gly-Gly-Gly-Gly)_y wherein y is 1, 2, 3, 4, 5, 6, 7, or 8. In other embodiments, additional amino acid sequences can be added to the N-

- 5 or C-terminus of the fusion protein to facilitate expression, steric flexibility, detection and/or isolation or purification.

A chimeric or fusion protein of the invention can be produced by standard recombinant DNA techniques. For example, DNA fragments coding for the different polypeptide sequences are ligated together in-frame in accordance with conventional 10 techniques, e.g., by employing blunt-ended or stagger-ended termini for ligation, restriction enzyme digestion to provide for appropriate termini, filling-in of cohesive ends as appropriate, alkaline phosphatase treatment to avoid undesirable joining, and enzymatic ligation. In another embodiment, the fusion gene can be synthesized by conventional techniques including automated DNA synthesizers. Alternatively, PCR 15 amplification of gene fragments can be carried out using anchor primers that give rise to complementary overhangs between two consecutive gene fragments that can subsequently be annealed and reamplified to generate a chimeric gene sequence (see, for example, Ausubel et al. (eds.) Current Protocols in Molecular Biology, John Wiley & Sons, 1992). Moreover, many expression vectors are commercially available that 20 encode a fusion moiety (e.g., an Fc region of an immunoglobulin heavy chain).

Immunoglobulin fusion polypeptide are known in the art and are described in e.g., U.S. Pat. Nos. 5,516,964; 5,225,538; 5,428,130; 5,514,582; 5,714,147; and 5,455,165.

Nucleic Acid Molecules, Constructs and Vectors

25 Exemplary embodiments of the instant invention feature engineered constructs designed to eliminate unwanted or undesirable by-products, in particular, unwanted or undesirable antibody (or immunoglobulin) by-products. In certain aspects, the constructs include components of naturally-occurring antibody gene sequences, wherein the components have been genetically altered, modified ,or engineered (e.g., 30 genetically engineered) such that the resultant construct expresses the desired protein (e.g., antibody) of interest in the absence of the unwanted or undesired by-product. Constructs can be generated using art-recognized techniques for producing recombinant nucleic acid molecules (e.g., comprising components of immunoglobulin chain genes) as described in detail below.

35 Antibody gene sequences encode antibodies of the various isotypes, including IgG (e.g., IgG1, IgG2, IgG3, IgG4), IgM, IgA1, IgA2, IgD, or IgE. Preferably, the antibody gene sequences encodes an antibody of the antibody is an IgG isotype. The

- 5 encoded immunoglobulin or antibody molecules can include full-length (*e.g.*, an IgG1 or IgG4 immunoglobulin) or alternatively can include only a fragment (*e.g.*, a Fc fragment).

It will be appreciated by the skilled artisan that nucleotide sequences encoding the antibodies of the instant invention can be derived from the nucleotide and amino acid sequences described in the present application or from additional sources of sequences of immunoglobulin genes known in the art using the genetic code and standard molecular biology techniques. The nucleic acid compositions of the present invention may be derived from known immunoglobulin DNA (*e.g.*, cDNA sequences). In particular, nucleotide sequences may be substantially identical to or derived from native V, D, J, or constant cDNA sequences. The sequences of heavy and light chain constant region genes are known in the art. Preferably, the constant region is human, but constant regions from other species, *e.g.*, rodent (*e.g.*, mouse or rat), primate (macaque), camel, or rabbit, can also be used. Constant regions from these species are known in the art (see *e.g.*, Kabat, E. A., et al. (1991) Sequences of Proteins of Immunological Interest, Fifth Edition, U.S. Department of Health and Human Services, NIH Publication No. 91-3242) and DNA fragments encompassing these regions can be obtained by standard PCR amplification. The heavy chain constant region can be an IgG1, IgG2, IgG3, IgG4, IgA, IgE, IgM or IgD constant region. Sequences for heavy chain constant regions are known in the art and can be found in, *e.g.*, NCBI NG_001019. In typical embodiments, the constant region is an IgG1 or IgG4 constant region. For an Fc fragment heavy chain gene, the Fc-encoding DNA can be operatively linked to a heavy chain leader sequence (*e.g.*, a heavy chain variable chain leader sequence) for direct expression.

Additional aspects of the invention include assembled immunoglobulin DNA cassette sequences. Assembled immunoglobulin cassette sequences include nucleotide sequences as well as amino acid sequences encoded by an immunoglobulin DNA cassette nucleotide sequence.

An exemplary human IgG1 constant region genomic sequence is hereby provided:

35 GTGAGTCCTGTCGACTCTAGAGCTTCTGGGCAGGCCAGGCCTGACTTTGGCTGG
GGCAGGGAGGGGGCTAACGGTGACGCAGGTGGCGCCAGGCCAGGCGCACACCCAATGCC
CATGAGCCCAGACACTGGACGCTGAACCTCGCGGACAGTTAACAGAACCCAGGGGCCTC
TGCGCCCTGGGCCAGCTCTGTCACACCCGCGGTACATGGCACCACCTCTTGC
AGCCTCCACCAAGGGCCATCGGTCTTCCCCCTGGCACCCCTCTCCAAGAGCACCTC
40 TGGGGGCACAGCGGCCCTGGCTGCCTGGTCAAGGACTACTTCCCCGAACCGGTGAC

5 GGTGTCTGGAACTCAGGCGCCCTGACCAGCGGCGTCACACCTCCGGCTGTCCT
 ACAGTCCTCAGGACTCTACTCCCTCAGCAGCGTGGTACCGTGCCCTCCAGCAGCTT
 GGGCACCCAGACCTACATCTGCAACGTGAATACAAGCCCAGCAACACCAAGGTGGA
 CAAGAAAGTTGGTGAGAGGCCAGCACAGGGAGGGAGGGTGTCTGCTGGAAGCCAGGC
 TCAGCGCTCTGCCCTGGACGCATCCCAGGCTATGCAGTCCCAGTCCAGGGCAGCAAGG
 CAGGCCCGTCTGCCCTTCACCCGGAGGCCTCTGCCCGCCCCACTCATGCTCAGGG
 AGAGGGTCTTCTGGCTTTTCCCCAGGCTCTGGCAGGCACAGGCTAGGTGCCCCATA
 ACCCAGGCCCTGCACACAAAGGGCAGGTGCTGGGCTCAGACCTGCCAAGAGGCCATA
 TCCGGGAGGACCCCTGCCCTGACCTAACGCCACCCAAAGGCCAAACTCTCCACTCC
 CTCAGCTCGGACACCTCTCTCCAGATTCCAGTAACCTCCAATCTCTCTCTG
 CAGAGCCCAAATCTTGACAAAAACTCACACATGCCACCGTGCCCAAGGTAAGGCCAG
 CCCAGGCCCTGCCCTCCAGCTCAAGGCGGGACAGGTGCCCCAGGTAAGCCTGCATCC
 AGGGACAGGCCAGGCCAGCCGGTGTGACACGTCCACCTCCATCTTCCCTCAGCACCT
 GAACCTCTGGGGGACCGTCAGTCTCCTCTCCCCAAAACCCAAGGACACCCTC
 ATGATCTCCGGACCCCTGAGGTACATGCGTGGTGGACGTGAGGTGACGACAGAAGACA
 CCTGAGGTCAAGTCAACTGGTACGTGGACGGCGTGGAGGTGACATGCAAGACA
 AAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGTGTGGTACCGTCCCTACCGTC
 CTGCACCAGGACTGGCTGAATGCAAGGAGTACAAGTCAAGGTCTCCAACAAAGCC
 CTCCCAGCCCCATCGAGAAAACCATCTCCAAGGCCAAAGGTGGACCCGTGGGTG
 CGAGGGCACATGGACAGAGGCCGGCTGGCCACCCCTCTGCCCTGAGAGTGACCGC
 TGTACCAACCTCTGTCCTACAGGGCAGCCCCGAGAACCAAGGTGTACACCCTGCC
 CCCATCCGGGAGGAGATGACCAAGAACCAAGGTGACCTGACCTGCCTGGTCAAAGG
 CTTCTATCCCAGCGACATGCCGTGGAGTGGAGAGCAATGGGCAGCCGGAGAACAA
 CTACAAGACCACGCCTCCCGTGTGGACTCCGACGGCTCTTCCCTATAGCAA
 GCTCACCGTGGACAAGAGCAGGTGGCAGCAGGGGAAAGTCTCTCATGCTCCGTGAT
 GCATGAGGTCTGACAACCACACGCAGAAGAGCCTCTCCCTGTCCCCGGTAA
 ATGA (SEQ ID NO:1)

An exemplary IgG4 constant region genomic sequence is hereby provided:

35 GTGAGTCCTGTCGACTCTAGAGCTTCTGGGGCAGGCCAGGCCTGACTTGGCTGGG
 GGCAGGGAGGGGCTAAGGTGACGCAAGGTGGCGCCAGCCAGGCGCACACCCAATGCC
 CATGAGCCCAGACACTGGACGCTGAACCTCGCGGACAGTTAACGACCCAGGGCCTC
 TGGCCCTGGCCAGCTCTGTCACACCGCGGTACATGGCACCAACCTCTTGC
 AGCCTCCACCAAGGGCCATCGGTCTTCCCCCTGGGCCCTGCTCCAGGAGCACCTC
 CGAGAGCACAGGCCCTGGCTGCCTGGTCAAGGACTACTTCCGAACCGGTGAC
 GGTGTCTGGAACTCAGGCCCTGACCAGCGCGTGCACACCTCCGGCTGTCCT
 ACAGTCCTCAGGACTCTACTCCCTCAGCAGCGTGGTACCGTGCCTCCAGCAGCTT
 GGGCACGAAGACCTACACCTGCAATGTAGATCACAAGGCCAGAACACCAAGGTGGA
 CAAGAGAGTTGGTGAGAGGCCAGCACAGGGAGGGAGGGTGTCTGCTGGAAGCCAGGC
 TCAGCCCTCCTGCCTGGACGCACCCGGCTGTGACGCCAGCCAGGGCAGCAAGG
 CAGGCCCATCTGTCCTCACCTGGAGGCCTGTGACCAACCCACTCATGCTCAGGG
 AGAGGGTCTCTGGATTTCCACCAAGGCTCCGGCAGGCCACAGGCTGGATGCCCT
 ACCCAGGCCCTGCCATACAGGGCAGGTGCTCGCCTCAGACCTGCCAAGAGCCAT
 ATCCGGGAGGACCCCTGCCCTGACCTAACGCCACCCAAAGGCCAAACTCTCCACTC
 CCTCAGCTCAGACACCTCTCCTCCAGATCTGAGTAACCTCCAATCTCTCT
 GCAGAGTCAAATATGGTCCCCATGCCACATGCCAGGTAAAGCAACCCAGGCC
 TCGCCCTCAGCTCAAGGCCAGGTGCCCCTAGAGTAGCCTGCATCCAGGGACAG
 GCCCGAGCCGGTGTGACGCATCCACCTCCATCTTCCCTCAGCACCTGAGTTCC
 GGGGGACCATCAGTCTTCTGTTCCCCAAAACCAAGGACACTCTCATGATCTC
 CCGGACCCCTGAGGTACGTGCGTGGTGGACGTGAGCCAGGAAGACCCAGGG
 CCAGTTCAACTGGTACGTGGATGGCGTGGAGGTGACATATGCCAAGACAAAGCCGCG
 GGAGGGAGCAGTTAACAGCACGTACCGTGTGGTACAGCTCCTCACCGTCTGCC
 GGACTGGCTGAACGCAAGGAGTACAAGTCAAGGTCTCCAACAAAGGCCCTCCGTC
 CTCCATCGAGAAAACCATCTCAAAGCCAAGGTGGACCCACGGGTGCGAGGGCC
 ACATGGACAGAGGTACGCTGCCACCCCTCTGCCCTGGGAGGTGACCGCTGTGCC
 CCTCTGTCCTACAGGGCAGCCCCGAGAGGCCACAGGTGTACACCCCTGCC
 AGGAGGGAGTGAACAAAGAACCAAGGTACGCCCTGACCTGCCAGGTCAAAGGCTTCT
 CCAGCGACATGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGAGAACAAACTACAAGA
 CCACGCCCTCCGTGTGGACTCCGACGGCTCTTCCCTACAGCAGGCTAACCG
 TGGACAAGAGCAGGTGGCAGGAGGGAAATGTCTCTCATGCTCCGTGATGCATGAGG

5 CTCTGCACAACCACTACACACAGAAGAGCCTCTCCCTGTCTGGGTAAATGA
(SEQ ID NO:3)

Antibody Production

10 Antibodies of the present invention are typically produced by recombinant expression. Nucleic acids encoding light and heavy chains can be inserted into expression vectors. The light and heavy chains can be cloned in the same or different expression vectors. The DNA segments encoding immunoglobulin chains are operably linked to control sequences in the expression vector(s) that ensure the expression of immunoglobulin polypeptides. Expression control sequences include,
15 but are not limited to, promoters (*e.g.*, naturally-associated or heterologous promoters), signal sequences, enhancer elements, and transcription termination sequences. Preferably, the expression control sequences are eukaryotic promoter systems in vectors capable of transforming or transfecting eukaryotic host cells (*e.g.*, COS or CHO cells).

20 Following manipulation of the isolated genetic material to provide polypeptides of the invention as set forth above, the genes are typically inserted in an expression vector for introduction into host cells that may be used to produce the desired quantity of modified antibody that, in turn, provides the claimed polypeptides. The term “vector” includes a nucleic acid construct often including a nucleic acid,
25 *e.g.*, a gene, and further including minimal elements necessary for nucleic acid replication, transcription, stability and/or protein expression or secretion from a host cell. Such constructs may exist as extrachromosomal elements or may be integrated into the genome of a host cell.

30 The term “expression vector” includes a specific type of vector wherein the nucleic acid construct is optimized for the high-level expression of a desired protein product. Expression vectors often have transcriptional regulatory agents, such as promoter and enhancer elements, optimized for high-levels of transcription in specific cell types and/ or optimized such that expression is constitutive based upon the use of a specific inducing agent. Expression vectors further have sequences that provide for
35 proper and/or enhanced translation of the protein. As known to those skilled in the art, such vectors may easily be selected from the group consisting of plasmids, phages, viruses, and retroviruses. The term “expression cassette” includes a nucleic acid

- 5 construct containing a gene and having elements in addition to the gene that allow for proper and or enhanced expression of that gene in a host cell.

The term "operably linked" includes a juxtaposition wherein the components are in a relationship permitting them to function in their intended manner (e.g., functionally linked). As an example, a promoter/enhancer operably linked to a
10 polynucleotide of interest is ligated to said polynucleotide such that expression of the polynucleotide of interest is achieved under conditions which activate expression directed by the promoter/enhancer. In regards to the invention described herein, operably linked also encompasses the relationship of splice donor and splice acceptor sites found in the primary transcript (pre-mRNA) of a gene of interest. Normally,
15 splice acceptor and donor sites are operably linked in that the two sequences are required and function together for splicing events to occur resulting in a mature messenger RNA.

Expression vectors are typically replicable in the host organisms either as episomes or as an integral part of the host chromosomal DNA. Commonly,
20 expression vectors contain selection markers (e.g., ampicillin-resistance, hygromycin-resistance, tetracycline resistance, kanamycin resistance or neomycin resistance) to permit detection of those cells transformed with the desired DNA sequences (see, e.g., Itakura *et al.*, U.S. Patent No. 4,704,362). In addition to the immunoglobulin DNA cassette sequences, insert sequences, and regulatory sequences, the recombinant
25 expression vectors of the invention may carry additional sequences, such as sequences that regulate replication of the vector in host cells (e.g., origins of replication) and selectable marker genes. The selectable marker gene facilitates selection of host cells into which the vector has been introduced (see e.g., U.S. Pat. Nos. 4,399,216, 4,634,665 and 5,179,017, all by Axel *et al.*). For example, typically the selectable
30 marker gene confers resistance to drugs, such as G418, hygromycin, or methotrexate, on a host cell into which the vector has been introduced. Preferred selectable marker genes include the dihydrofolate reductase (DHFR) gene (for use in dhfr⁻ host cells with methotrexate selection/amplification) and the neo gene (for G418 selection).

Once the vector has been incorporated into the appropriate host, the host is
35 maintained under conditions suitable for high level expression of the nucleotide sequences, and the collection and purification of the desired antibodies. Mammalian cells are preferred for expression and production of the antibodies of the present

5 invention. See, e.g., Winnacker, *From Genes to Clones*, VCH Publishers, N.Y., N.Y. (1987). Eukaryotic cells are preferred because a number of suitable host cell lines capable of secreting heterologous proteins (e.g., intact immunoglobulins) have been developed in the art, and include CHO cell lines, various COS cell lines, HeLa cells, preferably, myeloma cell lines, or transformed B-cells or hybridomas. Preferably, the
10 cells are non-human. Preferred mammalian host cells for expressing the antibodies of the invention include Chinese Hamster Ovary (CHO cells) (including dhfr⁻ CHO cells, described in Urlaub and Chasin (1980) *Proc. Natl. Acad. Sci. USA* 77:4216-4220, used with a DHFR selectable marker, e.g., as described in Kaufman and Sharp (1982) *Mol. Biol.* 159:601-621), lymphocytic cell lines, e.g., NSO myeloma cells and
15 SP2 cells, COS cells, and cells derived from a transgenic animal, e.g., mammary epithelial cell. Other suitable host cells are known to those skilled in the art.

Expression vectors for these cells can include expression control sequences, such as an origin of replication, a promoter, and an enhancer (Queen *et al.*, *Immunol. Rev.* 89:49 (1986)), and necessary processing information sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites, and transcriptional terminator sequences. Preferred expression control sequences are promoters derived from immunoglobulin genes, SV40, adenovirus, bovine papilloma virus, cytomegalovirus and the like. See, e.g., Co *et al.*, (1992) *J. Immunol.* 148:1149. Preferred regulatory sequences for mammalian host cell expression include viral elements that direct high levels of protein expression in mammalian cells, such as promoters and/or enhancers derived from FF-1a promoter and BGH poly A, cytomegalovirus (CMV) (such as the CMV promoter/ enhancer), Simian Virus 40 (SV40) (such as the SV40 promoter/enhancer), adenovirus (e.g., the adenovirus major late promoter (AdMLP)), and polyoma. For further description of viral regulatory elements, and sequences thereof, see, e.g., U.S. Pat. No. 5,168,062 by Stinski, U.S. Pat. No. 4,510,245 by Bell *et al.* and U.S. Patent No. 4,968,615 by Schaffner *et al.* In exemplary embodiments, the antibody heavy and light chain genes are operatively linked to enhancer/promoter regulatory elements (e.g., derived from SV40, CMV, adenovirus and the like, such as a CMV enhancer/AdMLP promoter regulatory element or an SV40 enhancer/AdMLP promoter regulatory element) to drive high levels of transcription of the genes. In exemplary embodiments of the invention, the construct include an internal ribosome entry site (IRES) to provide relatively high levels of polypeptides of the invention in

5 eukaryotic host cells. Compatible IRES sequences are disclosed in U.S. Pat. No. 6,193,980 that is also incorporated herein.

Alternatively, antibody-coding sequences can be incorporated in a transgene for introduction into the genome of a transgenic animal and subsequent expression in the milk of the transgenic animal (see, e.g., Deboer *et al.*, US 5,741,957, Rosen, US 10 5,304,489, and Meade *et al.*, US 5,849,992). Suitable transgenes include coding sequences for light and/or heavy chains in operable linkage with a promoter and enhancer from a mammary gland specific gene, such as casein or beta lactoglobulin.

Prokaryotic host cells may also be suitable for producing the antibodies of the invention. *E. coli* is one prokaryotic host particularly useful for cloning the 15 polynucleotides (e.g., DNA sequences) of the present invention. Other microbial hosts suitable for use include bacilli, such as *Bacillus subtilis*, enterobacteriaceae, such as *Escherichia*, *Salmonella*, and *Serratia*, and various *Pseudomonas* species. In these prokaryotic hosts, one can also make expression vectors, which will typically contain expression control sequences compatible with the host cell (e.g., an origin of 20 replication). In addition, any number of a variety of well-known promoters will be present, such as the lactose promoter system, a tryptophan (trp) promoter system, a beta-lactamase promoter system, or a promoter system from phage lambda. The promoters will typically control expression, optionally with an operator sequence, and have ribosome binding site sequences and the like, for initiating and completing 25 transcription and translation.

Expression of proteins in prokaryotes is most often carried out in *E. coli* with vectors containing constitutive or inducible promoters directing the expression of either fusion or non-fusion proteins. Fusion vectors add a number of amino acids to an antibody encoded therein, often to the constant region of the recombinant antibody, 30 without affecting specificity or antigen recognition of the antibody. Addition of the amino acids of the fusion peptide can add additional function to the antibody, for example as a marker (e.g., epitope tag such as myc or flag).

Other microbes, such as yeast, are also useful for expression. *Saccharomyces* is a preferred yeast host, with suitable vectors having expression control sequences 35 (e.g., promoters), an origin of replication, termination sequences, and the like as desired. Typical promoters include 3-phosphoglycerate kinase and other glycolytic enzymes. Inducible yeast promoters include, among others, promoters from alcohol

5 dehydrogenase, isocytochrome C, and enzymes responsible for maltose and galactose utilization.

Alternatively, antibodies of the invention can be produced in transgenic plants (e.g., tobacco, maize, soybean and alfalfa). Improved 'plantibody' vectors (Hendy *et al.* (1999) *J. Immunol. Methods* 231:137-146) and purification strategies coupled with 10 an increase in transformable crop species render such methods a practical and efficient means of producing recombinant immunoglobulins not only for human and animal therapy, but for industrial applications as well (e.g., catalytic antibodies). Moreover, plant produced antibodies have been shown to be safe and effective and avoid the use of animal-derived materials and therefore the risk of contamination with a transmissible 15 spongiform encephalopathy (TSE) agent. Further, the differences in glycosylation patterns of plant and mammalian cell-produced antibodies have little or no effect on antigen binding or specificity. In addition, no evidence of toxicity or HAMA has been observed in patients receiving topical oral application of a plant-derived secretory dimeric IgA antibody (see, e.g., Lerrick *et al.* (1998) *Res. Immunol.* 20 149:603-608).

Various methods may be used to express recombinant antibodies in transgenic plants. For example, antibody heavy and light chains can be independently cloned into expression vectors (e.g., *Agrobacterium tumefaciens* vectors), followed by the transformation of plant tissue *in vitro* with the recombinant bacterium or direct 25 transformation using, e.g., particles coated with the vector which are then physically introduced into the plant tissue using, e.g., ballistics. Subsequently, whole plants expressing individual chains are reconstituted followed by their sexual cross, ultimately resulting in the production of a fully assembled and functional antibody. Similar protocols have been used to express functional antibodies in tobacco plants (see, e.g., 30 Hiatt *et al.* (1989) *Nature* 342:76-87). In various embodiments, signal sequences may be utilized to promote the expression, binding and folding of unassembled antibody chains by directing the chains to the appropriate plant environment (e.g., the aqueous environment of the apoplasm or other specific plant tissues including tubers, fruit or seed) (see Fiedler *et al.* (1995) *Bio/Technology* 13:1090-1093). Plant bioreactors can 35 also be used to increase antibody yield and to significantly reduce costs.

Suitable host cells are discussed further in Goeddel (1990) Gene Expression Technology: Methods in Enzymology 185, Academic Press, San Diego, Calif.

5 Alternatively, the recombinant expression vector can be transcribed and translated *in vitro*, for example using T7 promoter regulatory sequences and T7 polymerase.

The vectors containing the polynucleotide sequences of interest (*e.g.*, the heavy and light chain encoding sequences and expression control sequences) can be transferred into the host cell by well-known methods, which vary depending on the 10 type of cellular host. For example, calcium chloride transfection is commonly utilized for prokaryotic cells, whereas calcium phosphate treatment, electroporation, lipofection, biolistics or viral-based transfection may be used for other cellular hosts. (See generally Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual* (Cold Spring Harbor Press, 2nd ed., 1989), incorporated by reference herein in its entirety 15 for all purposes.) Other methods used to transform mammalian cells include the use of polybrene, protoplast fusion, liposomes, electroporation, and microinjection (see generally, Sambrook *et al.*, *supra*). For production of transgenic animals, transgenes can be microinjected into fertilized oocytes, or can be incorporated into the genome of embryonic stem cells, and the nuclei of such cells transferred into enucleated oocytes.

20 When heavy and light chains are cloned on separate expression vectors, the vectors are co-transfected to obtain expression and assembly of intact immunoglobulins. Once expressed, the whole antibodies, their dimers, individual 25 light and heavy chains, or other immunoglobulin forms of the present invention can be purified according to standard procedures of the art, including ammonium sulfate precipitation, affinity columns, column chromatography, HPLC purification, gel electrophoresis and the like (see generally Scopes, *Protein Purification* (Springer-Verlag, N.Y., (1982)). Substantially pure immunoglobulins of at least about 90 to 95% homogeneity are preferred, and 98 to 99% or more homogeneity most preferred, for pharmaceutical uses.

30 An immunoglobulin or antibody produced according to the instant invention molecule can be derivatized or linked to another functional molecule (*e.g.*, another peptide or protein). Accordingly, the antibodies and antibody portions or otherwise modified forms of the antibodies of the invention described herein, may be further derivatized for use in research, diagnostic and/or therapeutic contexts. For example, 35 an antibody or antibody portion of the invention can be functionally linked (by chemical coupling, genetic fusion, noncovalent association or otherwise) to one or more other molecular entities, such as another antibody (*e.g.*, a bispecific antibody or

5 a diabody), a detectable agent, a cytotoxic agent, a pharmaceutical agent, and/or a protein or peptide that can mediate associate of the antibody or antibody portion with another molecule (such as a streptavidin core region or a polyhistidine tag).

One type of derivatized antibody is produced by crosslinking two or more antibodies (of the same type or of different types, *e.g.*, to create bispecific antibodies).

10 Suitable crosslinkers include those that are heterobifunctional, having two distinctly reactive groups separated by an appropriate spacer (*e.g.*, m-maleimidobenzoyl-N-hydroxysuccinimide ester) or homobifunctional (*e.g.*, disuccinimidyl suberate). Such linkers are available from Pierce Chemical Company, Rockford, IL.

15 Exemplary fluorescent detectable agents include fluorescein, fluorescein isothiocyanate, rhodamine, 5-dimethylamine-1-naphthalenesulfon-yl chloride, phycoerythrin and the like. An antibody may also be derivatized with detectable enzymes, such as alkaline phosphatase, horseradish peroxidase, P-galactosidase, acetylcholinesterase, glucose oxidase and the like. When an antibody is derivatized with a detectable enzyme, it is detected by adding additional reagents that the enzyme uses to produce a detectable reaction product. For example, when the detectable agent horseradish peroxidase is present, the addition of hydrogen peroxide and diaminobenzidine leads to a colored reaction product, which is detectable. An antibody may also be derivatized with a prosthetic group (*e.g.*, streptavidin/biotin and avidin/biotin). For example, an antibody may be derivatized with biotin, and detected through indirect measurement of avidin or streptavidin binding. Examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; examples of bioluminescent materials include luciferase, luciferin, and aequorin, and examples of suitable radioactive material include ^{125}I , ^{131}I , ^{35}S or ^3H . An antibody (or fragment thereof) may also be conjugated to a therapeutic moiety such as a cytotoxin or other therapeutic protein. Alternatively, an antibody can be conjugated to a second antibody to form an antibody heteroconjugate as described by Segal in U.S. Patent No. 4,676,980.

5 Expression Vectors for Decreasing or Eliminating Unwanted Polypeptide By-Products

During the development of a protein expression system for therapeutic proteins, HPLC analysis of purified target product identified unexpected low molecular weight (LMW) species of peptides. More specifically, undesired 10 polypeptide by-products were observed in a CHO (Chinese hamster ovary) cell line developed to express the 3D6 antibody. This antibody has been described elsewhere and is the result of efforts to develop an immunotherapeutic agent useful for the treatment of Alzheimer's disease. It has specificity for the A-beta peptide and has been demonstrated to be efficacious in clearing A-beta plaques. The CHO cell line 15 was developed using art accepted methods and contained copies of the heavy and light chain of the 3D6 antibody in addition to genes for selective culture of expression cassette containing cells.

Examination of a number of clonal isolates of the cell line demonstrated that 20 production of the LMW species was not a phenomenon specific to the clone being utilized, *i.e.*, a minor fraction of the total protein produced in all of the cell lines tested was of the unexpected LMW species. It was further observed that the fraction of LMW species relative to total protein increased when protein expression was induced. Further evaluation of the polypeptides using mass spectrometry indicated that the 25 LMW species contained amino acids not predicted by the exonic sequences of the gene.

The top panel of Figure 1 schematically presents the 3D6 heavy chain expression cassette showing the relation of introns and exons as well as the position of the internal ribosomal entry site (IRES) and dihydrofolate reductase (DHFR) selectable marker gene. The exons shown are variable heavy (V_{H1}), hinge and 30 constant heavy 1, 2 and 3 (C_{H1} , C_{H2} , C_{H3}). The introns of the expression cassette are denoted Int1, Int2, Int3 and Int4. Figure 1 further illustrates the predicted correct splicing events for the mRNA derived from the expression cassette. The middle panel shows the correctly spliced mRNA containing only intronic sequences of the bicistronic transcript.

35 Scrutiny of the intronic and exonic sequences in the expression vector and mass spectrometry data pointed to RNA polymerase intron read-through (IRT) of a specific splice site junction. Since the organization of the introns and exons and

5 splice site donor and acceptor sites contained in the expression vector were substantially identical to those as they existed in the original genomic form of the gene, the missplicing event was not predictable.

The bottom panel of Figure 1 illustrates the predicted product generated by intron read-through of the fourth intron. Figure 2 provides sequence information
10 showing the sense and anti-sense strands of the DNA sequence in the region of the fourth intron of the genomic sequence of the 3D6 antibody expression vector. The splice junctions (splice donor and acceptor sites) are denoted by vertical lines perpendicular to the nucleic acid sequence. DNA corresponding to intronic sequence is shown underlined and in italics. Predicted amino acids for desired and read-
15 through by-product polypeptides are shown below the anti-sense strand of the genomic DNA. The amino acid sequence of polypeptide derived from correctly spliced RNA is shown in bold uppercase lettering; polypeptide by-products derived from incorrectly spliced RNA is shown in lowercase font.

The present invention describes materials and methods for designing protein
20 expression cassettes and vectors such that intron read-through (IRT) and unwanted polypeptide byproducts are substantially reduced or eliminated entirely. In part, the invention provides on the novel design of vectors wherein the natural operative association of introns and exons in an isolated nucleic acid coding for a protein of interest are altered such that IRT is reduced or eliminated thereby reducing or
25 eliminating unwanted IRT polypeptide species. The unique alterations are particularly suitable for IgG1 or IgG4 antibodies, but may be used for any gene of interest. Moreover the vectors of the instant invention having introns and exons with altered natural operative associations demonstrate not only reduced or eliminated IRT by-products but also increased protein expression levels relative to vectors designed
30 using standard art recognized techniques.

Examples

Materials and Methods

Throughout the examples, materials and methods as exemplified in the
35 following texts were used unless otherwise stated:

In general, the practice of the present invention employs art-recognized techniques in molecular biology, recombinant DNA technology, and immunology

5 especially, e.g. antibody technology. See, e.g., Sambrook, Fritsch and Maniatis, Molecular Cloning: Cold Spring Harbor laboratory Press (1989); Antibody Engineering Protocols (Methods in Molecular Biology), 510, Paul, S., Human Press (1996); Antibody Engineering: A Practical Approach (Practical Approach Series, 169), McCafferty, Ed IRL Press (1996); Antibodies: A Laboratory Manual, Harlow *et al* Cold Spring Harbor Press, (1999); and Current Protocols in Molecular Biology eds. Ausubel *et al* John Wiley & Sons (1992).

Example 1. Quantification of Intron Read-Through Transcription

In order to quantify the relative amount of aberrant transcript formed due to 15 intron read through, a quantitative PCR assay was designed. The approach for evaluating IRT transcription is graphically outlined in Figure 3. Specifically, a quantitative PCR assay was devised using a TaqMan™ system, in which PCR amplification was employed to quantitate nucleic acid species of interest. Three probe-primer sets were designed to determine the fraction of intron read-through 20 mRNA being produced. The first probe-primer set was designed to quantitate the level of transcription of sequence of an exon in natural operative association with an intron of interest. In the case of the 3D6 heavy chain expression cassette, mRNA species containing the 3D6 second constant heavy chain (CH2) exon was targeted. This provided a measure of total 3D6 mRNA production. The second probe primer 25 set bridged the intron and exon in operative association, here the CH2 exon - fourth intron interface of the 3D6 expression cassette. Amplification derived from this probe primer set indicated the presence of intron read-through transcript containing the 5' splice donor sequence as well as sequence bridging the CH2 exon and intron 4. The third probe-primer set targeted sequence of the fourth intron. This probe set provided 30 quantification of the fraction of incorrectly spliced RNA comprising internal intron 4 sequence.

Figure 4 shows the results of the Q-PCR assay using the probe primer sets as described. Briefly, CHO cells containing the stably integrated expression vector were seeded and maintained in culture for two weeks. At day seven the cultures were 35 induced to increase protein expression. During the course of the experiment, samples of the cell culture were lysed and RNA content evaluated in assays using probe and primer sets specific for the CH2 exon or specific for intron as described in the

5 preceding paragraph. The chart demonstrates a low level of incorrectly spliced RNA product prior to induction and an increasing percentage of intron 4 containing RNA over time post-induction. This method of Q-PCR described here predicts the likelihood that a particular expression cassette containing introns and exons in naturally operative association will yield intron read through by-products .

10 While details for quantifying IRT of the 3D6 antibody expression system are explicitly provided, the technique can be implemented in any protein expression system wherein the potential of IRT exists. This novel approach is, therefore, especially useful for evaluating whether the vectors of this invention (described in detail below) should be adopted for a particular protein of interest such that the
15 production of unwanted IRT polypeptide by-products are avoided. When IRT transcription is in an abundance of greater than about 0.1% - 1%, vectors employing altered natural operative association can be employed to express the desired protein. It will be readily apparent to one of skill in the art that the methods for detecting
20 intron-read through mRNA and, hence, predicting intron read-through polypeptides is applicable to any protein expression system wherein splicing events occur. For example the system may be used with any eukaryotic cell system, e.g.
Saccharomyces, Drosophila, mouse, monkey, rabbit, rat, or human cell based
systems.

25 Example 2. Vectors with Introns and Exons Having Modified Natural Operative Association

Expression vectors were devised wherein the natural operative association of the introns and exons were modified. Two exemplary vectors sequences are shown in Figure 5. This figure illustrates expression constructs developed to resolve the
30 problem of intron read-through by-products. The top panel graphically depicts the genomic, intronic – exonic, organization of a generic antibody heavy chain containing the exons for a variable region (V_H), three constant regions (C_{H1}, C_{H2}, C_{H3}) and a hinge region. The middle and bottom drawings describe modifications to the genomic sequence incorporated into expression vectors which eliminated intron read through
35 heavy chain by-products.

CHO cells expressing the 3D6 light chain were transformed with either the complete genomic heavy chain sequence of the 3D6 antibody or transformed with

5 modified 3D6 heavy chain expression vectors wherein the natural operative
association of introns and exons were modified. The cells were cultured using
standard techniques for the purpose of protein expression as described in the Materials
and Methods. Antibodies were purified from conditioned supernatant and
subsequently fractionated using denaturing reverse phase (RP) HPLC (Figure 6). The
10 columns were run such that heavy and light chain constituents of the antibody were
resolved.

In the top trace, representing the fractionation of a 3D6 genomic clone protein
preparation, the heavy and light chains peaks are readily apparent. In addition, a
small peak can be discerned fractionating between the heavy and light chain
15 corresponding to heavy chain intron read-through product.

The bottom trace is an example of an expression system in which the problem
of intron read-through has been reduced. As in the top trace, light chain and heavy
chain peaks are clearly present, however, the level of IRT has been reduced to below
the limit of detection. The finding has been extended to other vectors in which the
20 natural operative association of exons and introns have been altered. For example, the
HCΔIntron 4 sequence described in Figure 5 similarly reduces IRT to undetectable
levels.

Example 3. Origin and Description of the Anti-5T4 Coding Sequence

25 Anti-5T4 H8 was obtained as a mouse monoclonal antibody against soluble
5T4. The anti-5T4 H8 antibody was humanized by CDR grafting (VH, DP75
germline framework; VL DPK24 germline framework) and the variable regions were
subcloned into vectors containing human IgG4 heavy chain constant domain (for VH
domain) or human kappa light chain constant domain (VL domain) as appropriate.
30 The humanized antibody is referred to as huH8. The hinge stabilizing mutation, Ser
241 to Pro, was introduced into the human IgG4. Sequences encoding huH8 heavy
and light chain were ligated into expression vectors driven by the mouse CMV
enhancer/promoter and containing the selectable marker genes dihydrofolate
reductase (heavy chain expression vector) or neomycin resistance (light chain
35 expression vector).

5 Example 4. Intron Removal Increases Protein Expression

To determine the effect of intron removal on antibody expression, expression constructs of several antibodies were created with differing numbers of introns. Variable regions of three different antibodies, 12A11v3.1, 356A11 and huH8, were each stably expressed in CHO cells with three constant region expression constructs
10 containing genomic sequence, cDNA sequence, and genomic sequence with three introns deleted (i.e., intron between CH1 and hinge region, intron between the hinge region and CH2, and intron between CH2 and CH3). For constructs that gave poor expression, such as 12A11v3.1 and huH8, removal of the three introns or all introns, i.e., cDNA, gave a significant increase in antibody expression. More specifically,
15 about a five-fold increase in expression was detected in the three-intron deleted construct for 12A11v3.1 compared to the genomic sequence. The 12A11v3.1 construct having the cDNA sequence showed over a six-fold increase in expression relative to the genomic sequence. The increased expression in CHO cells for huH8 antibodies was almost four-fold and nine-fold for the three-intron deleted and the
20 cDNA construct, respectively, compared to the genomic sequence. Typically, well expressed antibodies did not show a significant change in CHO-cell expression between the intron-deleted sequences and the genomic sequences.

Although the foregoing invention has been described in detail for purposes of clarity of understanding, it will be obvious that certain modifications may be practiced within the scope of the appended claims. All publications and patent documents cited herein, as well as text appearing in the figures and sequence listing, are hereby incorporated by reference in their entirety for all purposes to the same extent as if each were so individually denoted.

5 We claim:

1. A nucleic acid molecule comprising a nucleotide sequence having one or more intron and exon sequences, wherein at least one intron sequence is deleted compared to the naturally-occurring genomic sequence to reduce a mis-spliced or an intron read-through (IRT) by-product.

10

2. A nucleic acid molecule that comprises a nucleotide sequence comprising one or more intron and exon sequences, wherein at least one intron sequence is deleted compared to the naturally-occurring genomic sequence to enhance protein expression.

15

3. The nucleic acid of claim 2, wherein at least three intron sequences are deleted.

20 4. The nucleic acid molecule of any of claims 1-3, wherein the nucleotide sequence encodes an antibody heavy chain or a fragment thereof.

25 5. The nucleic acid molecule of claim 4, wherein the antibody heavy chain or fragment thereof comprises a heavy chain variable region, a hinge region, a first constant region (C_{H1}), a second constant region (C_{H2}), and third constant region (C_{H3}) of a human immunoglobulin G subtype.

6. The nucleic acid molecule of claim 5, wherein the immunoglobulin G subtype is a human IgG1 or human IgG4.

30 7. The nucleic acid molecule of claim 6, wherein the human IgG1 or human IgG4 is mutated.

35 8. The nucleic acid molecule of claim 5, wherein an intron between the C_{H2} region and the C_{H3} region of the immunoglobulin heavy chain constant region is deleted.

5 9. The nucleic acid molecule of claim 8, further comprising a deletion of an
intron between the C_H1 region and the hinge region.

10 10. The nucleic acid molecule of claim 8, further comprising a deletion of an
intron between the hinge region and the C_H2 region.

10 11. The nucleic acid molecule of claim 5, having a heavy chain that comprises
one intron between the heavy chain variable region and the C_H1 region.

15 12. The nucleic acid molecule of claim 5, wherein the nucleotide sequence
encoding the heavy chain hinge region, and the first, second and third constant
regions comprises a sequence at least 95% identical to the nucleotide sequence shown
in Figure 8 (SEQ ID NO:1).

20 13. The nucleic acid molecule of claim 5, wherein the nucleotide sequence
encoding the heavy chain hinge region, and a first, second, and third constant region
comprises a sequence at least 95% identical to the nucleotide sequence shown in
Figure 9 (SEQ ID NO:3).

25 14. The nucleic acid molecule of claim 8, wherein the deletion of the intron
between CH2 and CH3 corresponds to about nucleotides 1409 to 1505 of human IgG1
as shown in Figure 8 (SEQ ID NO:1).

30 15. The nucleic acid molecule of claim 8, wherein the deletion of the intron
between CH2 and CH3 corresponds to about nucleotides 1401 to 1497 of human IgG4
as shown in Figure 9 (SEQ ID NO:3).

35 16. The nucleic acid molecule of claim 9, wherein the deletion of the intron
between CH1 and the hinge region corresponds to about nucleotides 525 to 915 of
human IgG1 as shown in Figure 8 (SEQ ID NO:1).

35 17. The nucleic acid molecule of claim 9, wherein the deletion of the intron
between CH1 and the hinge region corresponds to about nucleotides 525 to 916 of
human IgG4 as shown in Figure 9 (SEQ ID NO:3).

5

18. The nucleic acid molecule of claim 10, wherein the deletion of the intron between the hinge region and C_H2 corresponds to about nucleotides 961 to 1078 of human IgG1 as shown in Figure 8 (SEQ ID NO:1).

10 19. The nucleic acid molecule of claim 10, wherein the deletion of the intron between the hinge region and C_H2 corresponds to about nucleotides 953 to 1070 of human IgG4 as shown in Figure 9 (SEQ ID NO:3).

15 20. A nucleic acid molecule comprising a nucleotide sequence encoding human IgG1, wherein said nucleotide sequence is at least 90% identical to the sequence shown in Figure 10 (SEQ ID NO:5).

20 21. A nucleic acid molecule comprising a nucleotide sequence encoding human IgG4, wherein said nucleotide sequence is at least 90% identical to the sequence shown in Figure 11 (SEQ ID NO:6).

25 22. A genomic nucleotide sequence encoding a human heavy chain constant region, or a mutated form thereof, wherein said nucleotide sequence lacks at least one intron present in the naturally-occurring genomic sequence, and wherein said intron facilitates intron-read through.

30 23. A genomic nucleotide sequence encoding a human IgG1, or a mutated form thereof, wherein said nucleotide sequence lacks at least one intron present in the naturally-occurring genomic sequence, and wherein said intron facilitates intron-read through.

24. The nucleotide sequence of either of claims 22 or 23, wherein the at least one intron is the intron between C_H2 and C_H3 of the constant region.

35 25. A genomic nucleotide sequence encoding a human IgG4, or a mutated form thereof, wherein said genomic sequence lacks three introns present in the naturally-occurring genomic sequence.

5

26. The nucleotide sequence of claim 25, wherein the introns are the inton between C_H1 and hinge region, the inton between the hinge region and C_H2, and the inton between C_H2 and C_H3.

10 27. A nucleic acid molecule comprising a nucleotide sequence represented by the formula:

V_H -Int1-C_H1-Int2-Hinge-Int3-C_H2-C_H3,

wherein V_H is a nucleotide sequence encoding a heavy chain variable region;

C_H1, C_H2, and C_H3 are nucleotide sequences encoding the corresponding

15 heavy chain constant region;

Hinge is a nucleotide sequence encoding a hinge region of a heavy chain constant region; and

Int1, Int2 and Int3 are introns from the heavy chain genomic sequence.

20 28. The nucleic acid molecule of claim 27, wherein the nucleotide sequence encodes a human immunoglobulin G heavy chain.

29. A nucleic acid molecule comprising a nucleotide sequence represented by the formula:

25 V_H -Int1-C_H1-Hinge-C_H2-C_H3,

wherein V_H is a nucleotide sequence encoding a heavy chain variable region;

C_H1, C_H2, and C_H3 are nucleotide sequences encoding the corresponding

heavy chain constant region;

Hinge is a nucleotide sequence encoding a hinge region of a heavy chain

30 constant region; and

Int1 is an inton from the heavy chain genomic sequence.

30. The nucleic acid molecule of claim 29, wherein the nucleotide sequence encodes a human immunoglobulin G heavy chain.

35

31. An expression cassette comprising the nucleic acid molecule of claim 5.

5 32. An expression vector comprising the nucleic acid molecule of claim 5.

 33. The expression vector of claim 32, further comprising one or more nucleotide sequences that enhance replication, selection, mRNA transcription, mRNA stability, protein expression or protein secretion in a host cell.

10 34. A host cell comprising the nucleic acid molecule of claim 5.

 35. A host cell comprising the expression cassette of claim 31.

15 36. A host cell comprising the expression vector of claim 32.

 37. The host cell of claim 36, which is a Chinese Hamster Ovary (CHO) cell.

 38. A method of expressing a recombinant antibody or fragment thereof substantially free of an intron read-through (IRT) product, comprising:
20 introducing the nucleic acid molecule of claim 5 into a mammalian host cell;
 culturing said host cell under conditions that allow expression of the recombinant antibody or fragment thereof, thereby producing a culture of host cells;
 and
25 obtaining the recombinant antibody or fragment thereof from the culture of host cells.

 39. The method of claim 38, further comprising the step of identifying an IRT product in a nucleic acid sample from the host cell.

30 40. The method of claim 39, wherein the identification step comprises:
 obtaining a nucleic acid sample from the culture of host cells;
 contacting said nucleic acid sample with nucleic acid probes complementary to an intron and adjacent exon sequence, under conditions that allow hybridization
35 between the nucleic acid sample and the probes;

5 detecting the resulting complex, wherein detection in said sample of a complex, using the nucleic acid probe complementary to the intron sequence is indicative of the presence of the IRT product.

10 41. The method of claim 38, wherein said host cell comprises a nucleotide sequence encoding a light chain variable region and a constant region.

15 42. A method for enhancing expression of a recombinant antibody or fragment thereof, comprising:

introducing the nucleic acid molecule of claim 5 into a mammalian host cell;
culturing said host cell under conditions that allow expression of the recombinant antibody, thereby producing a culture of host cells; and
obtaining the recombinant antibody from the culture of host cells.

20 43. The method of claim 42, wherein said host cell comprises a nucleotide sequence encoding a light chain variable region and a constant region.

25 44. A method for producing a recombinant antibody or fragment thereof substantially devoid of intron read-through (IRT) heavy chain by-product, comprising:

culturing a mammalian host cell comprising the nucleic acid molecule of claim 5 and a nucleic acid encoding an antibody light chain, under conditions such that the heavy and light chains are expressed.

30 45. The method of claim 44, further comprising purifying the heavy and light chains from the culture.

46. A method for enhancing expression of a recombinant antibody or fragment thereof, comprising:

35 culturing a mammalian host cell comprising the nucleic acid molecule of claim 5, and a nucleic acid encoding an antibody light chain, under conditions such that the heavy and light chains are expressed.

5 47. The method of claim 46, further comprising purifying the heavy and light
chains from the culture.

48. A method for detecting an IRT product, in a sample, comprising:

obtaining a nucleic acid sample from a recombinant cell;

10 contacting said nucleic acid sample with nucleic acid probes complementary
to an intron and adjacent exon sequence, under conditions that allow hybridization of
the nucleic acid sample and the probes;

detecting the resulting complex, wherein detection in said sample of a
complex, using the nucleic acid probe complementary to the intron sequence is
15 indicative of the presence of the IRT product.

49. An antibody, or antigen-binding fragment thereof, made by the method
comprising the steps of either of claims 38 or 40 under suitable conditions to allow
expression and assembly of the antibody or fragment thereof.

20 50. The antibody of claim 49, which is a chimeric, humanized, CDR-grafted or
an *in vitro* generated antibody.

51. The antibody of claim 50, which is a humanized antibody.

25 52. The antibody of claim 51, which binds to human 5T4.

53. A pharmaceutical composition comprising the antibody of claim 49, and a
pharmaceutically acceptable carrier.

30

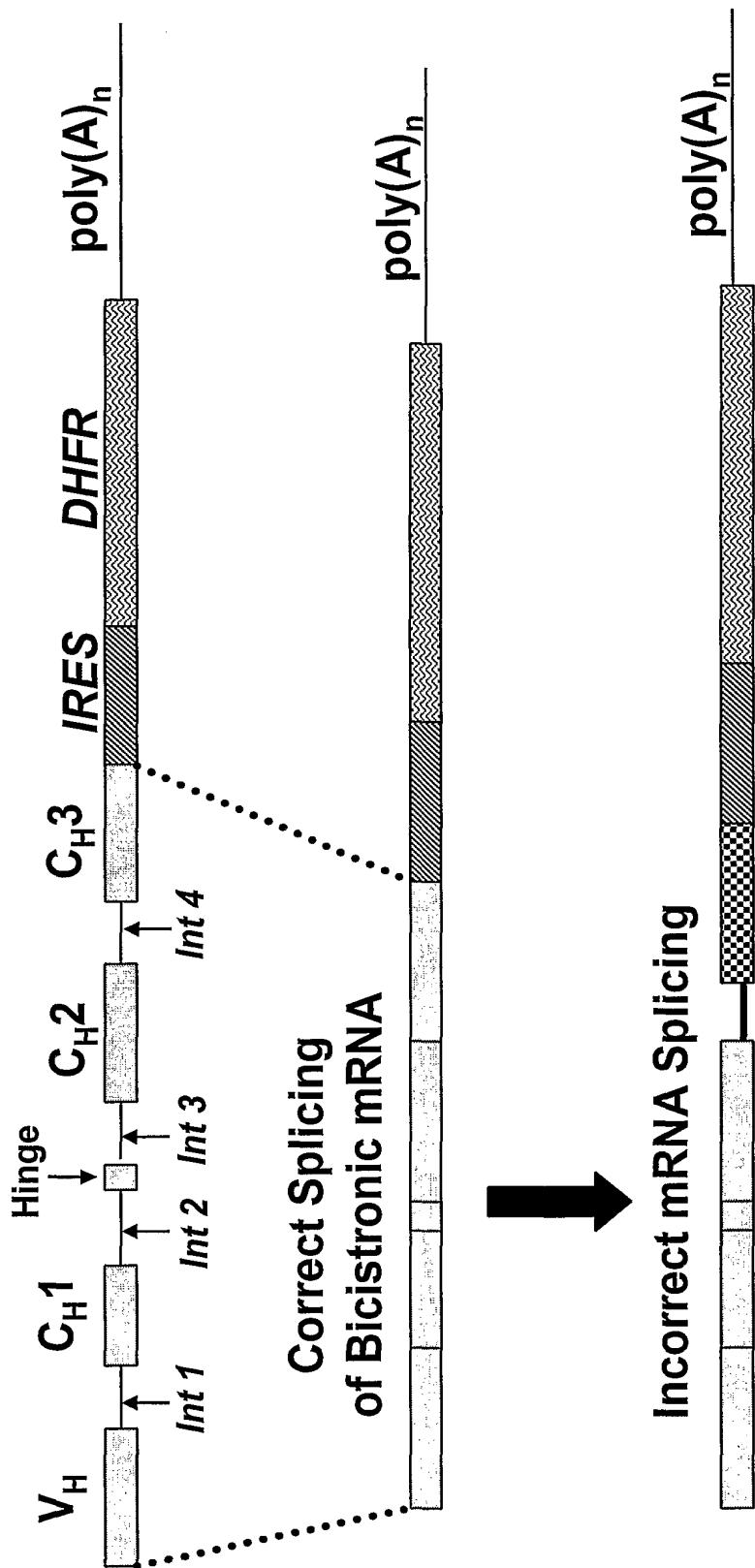
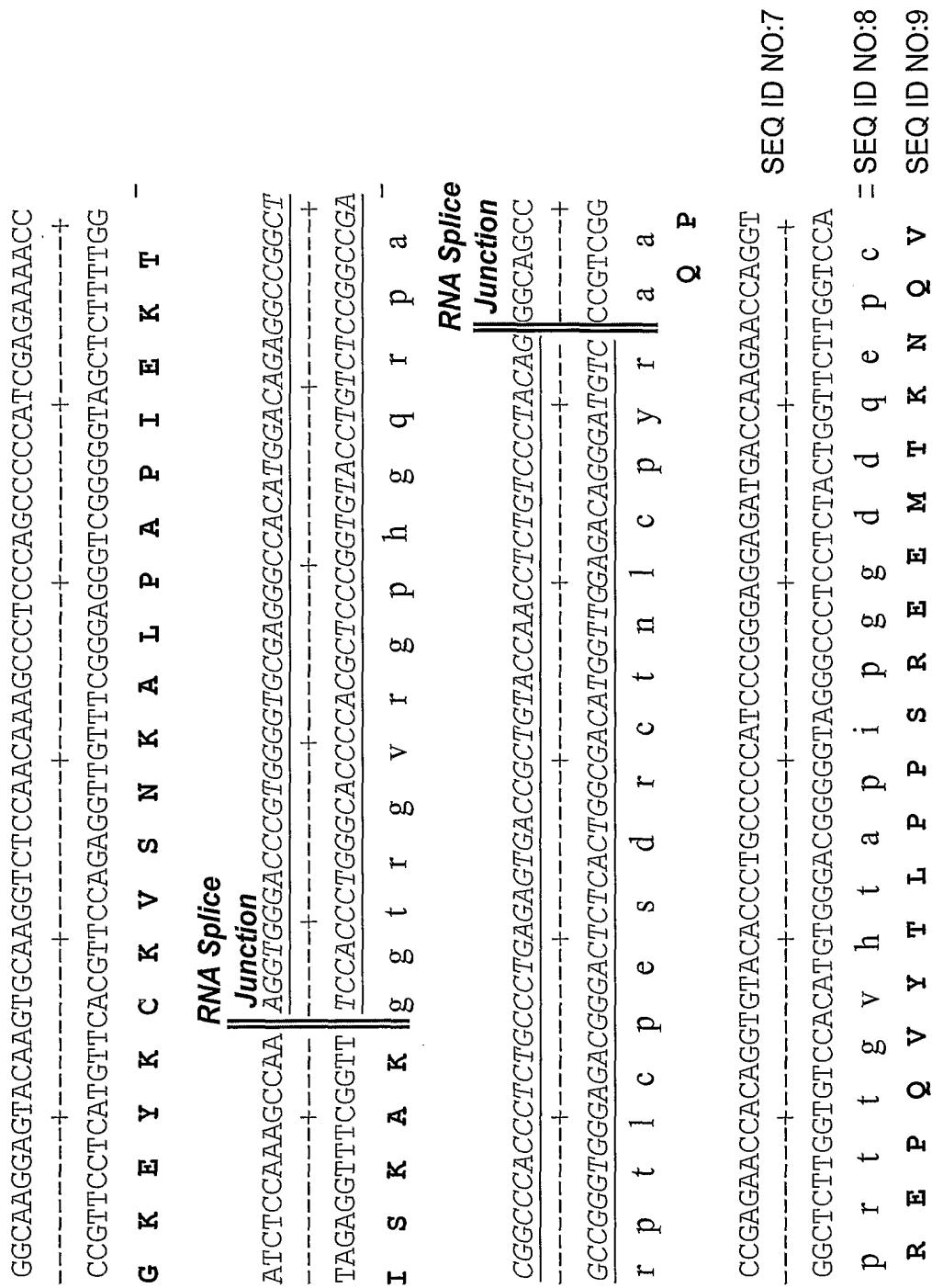


Figure 1

**Figure 2**

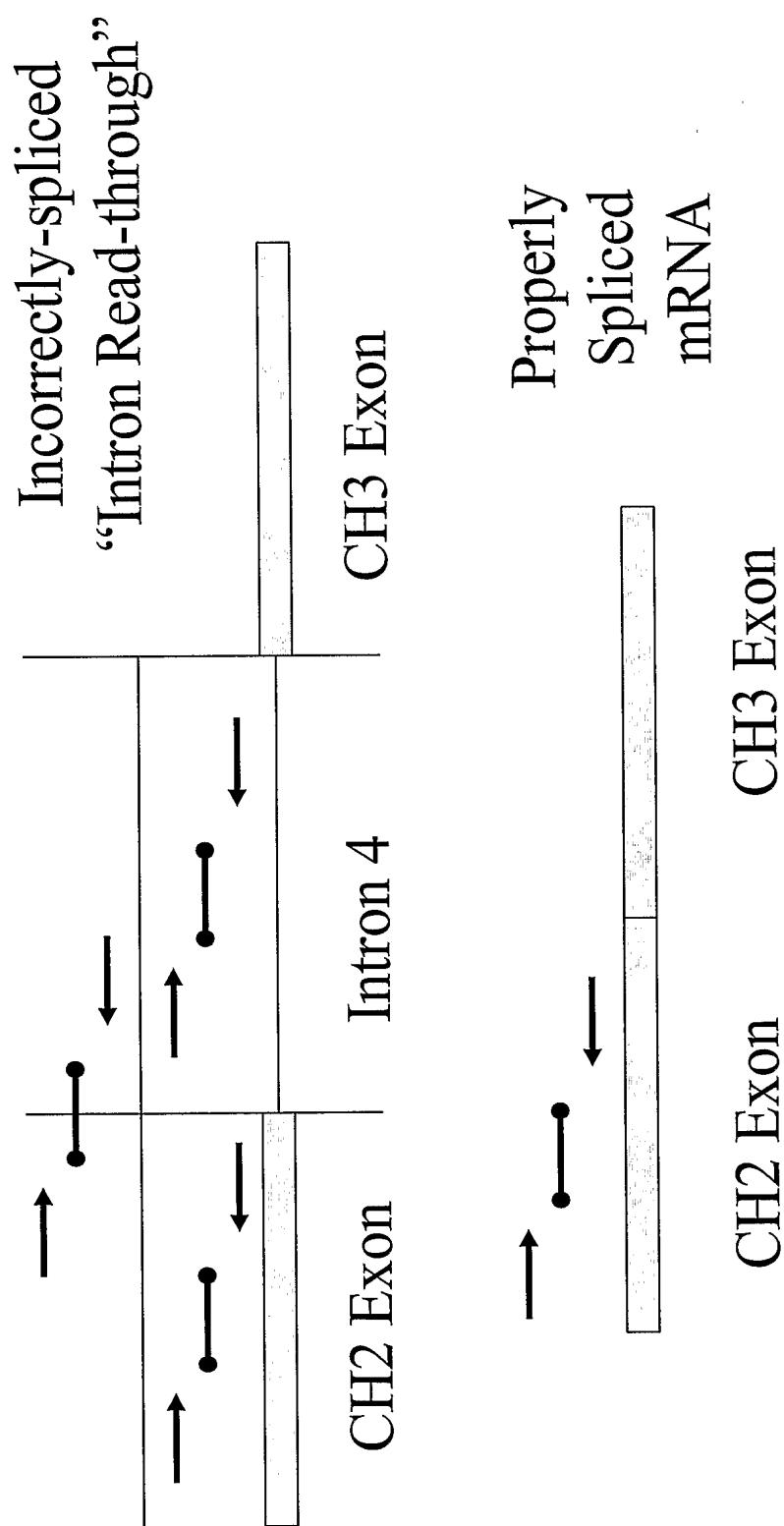


Figure 3

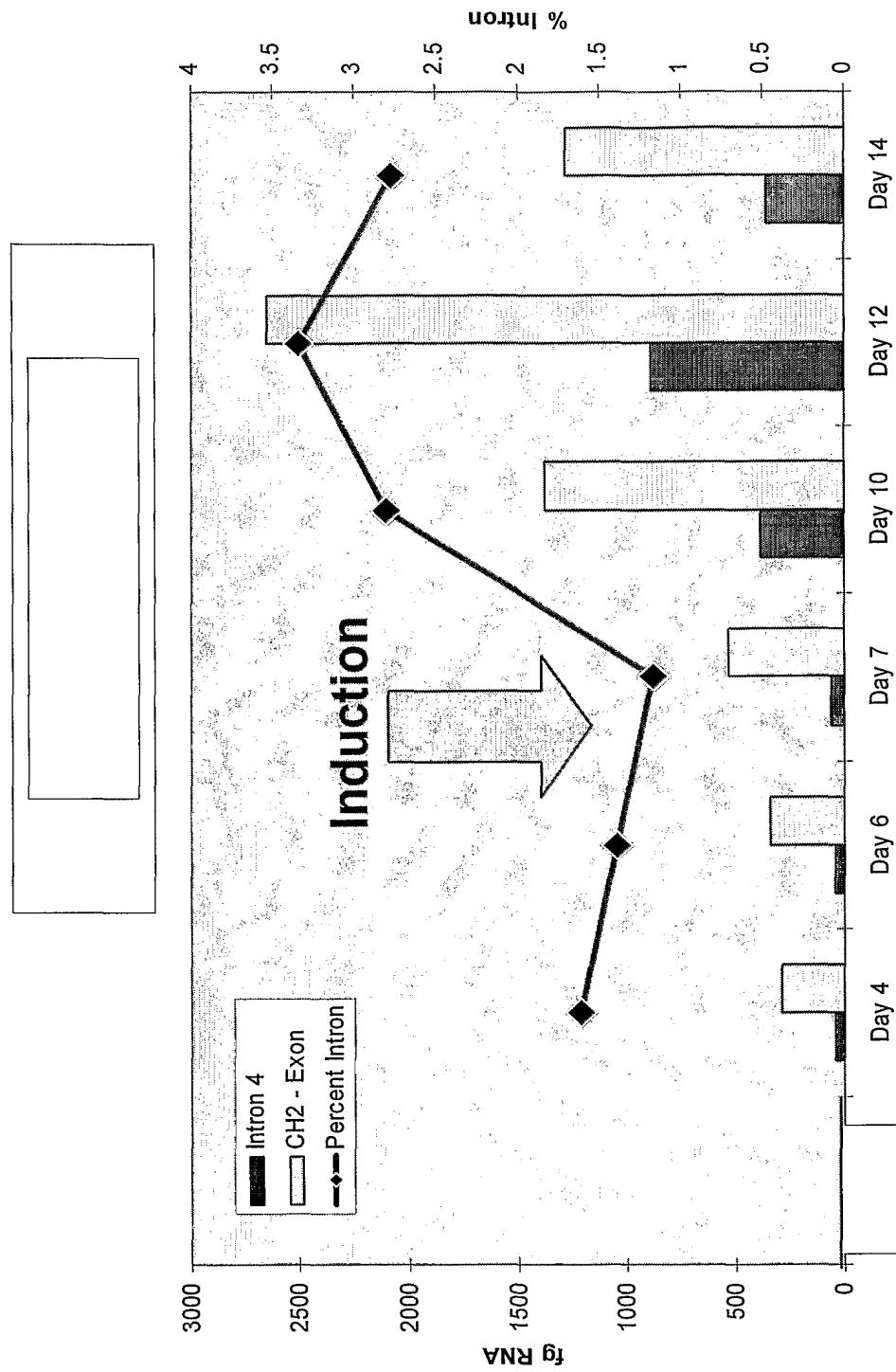


Figure 4

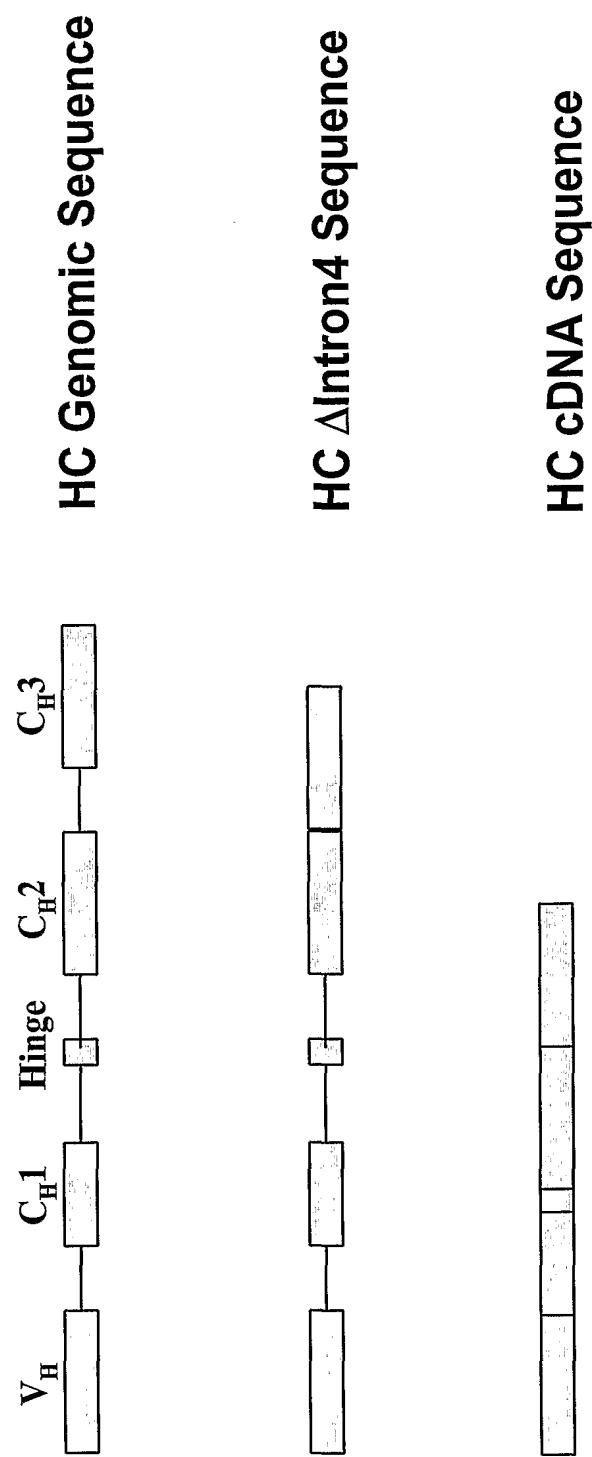


Figure 5

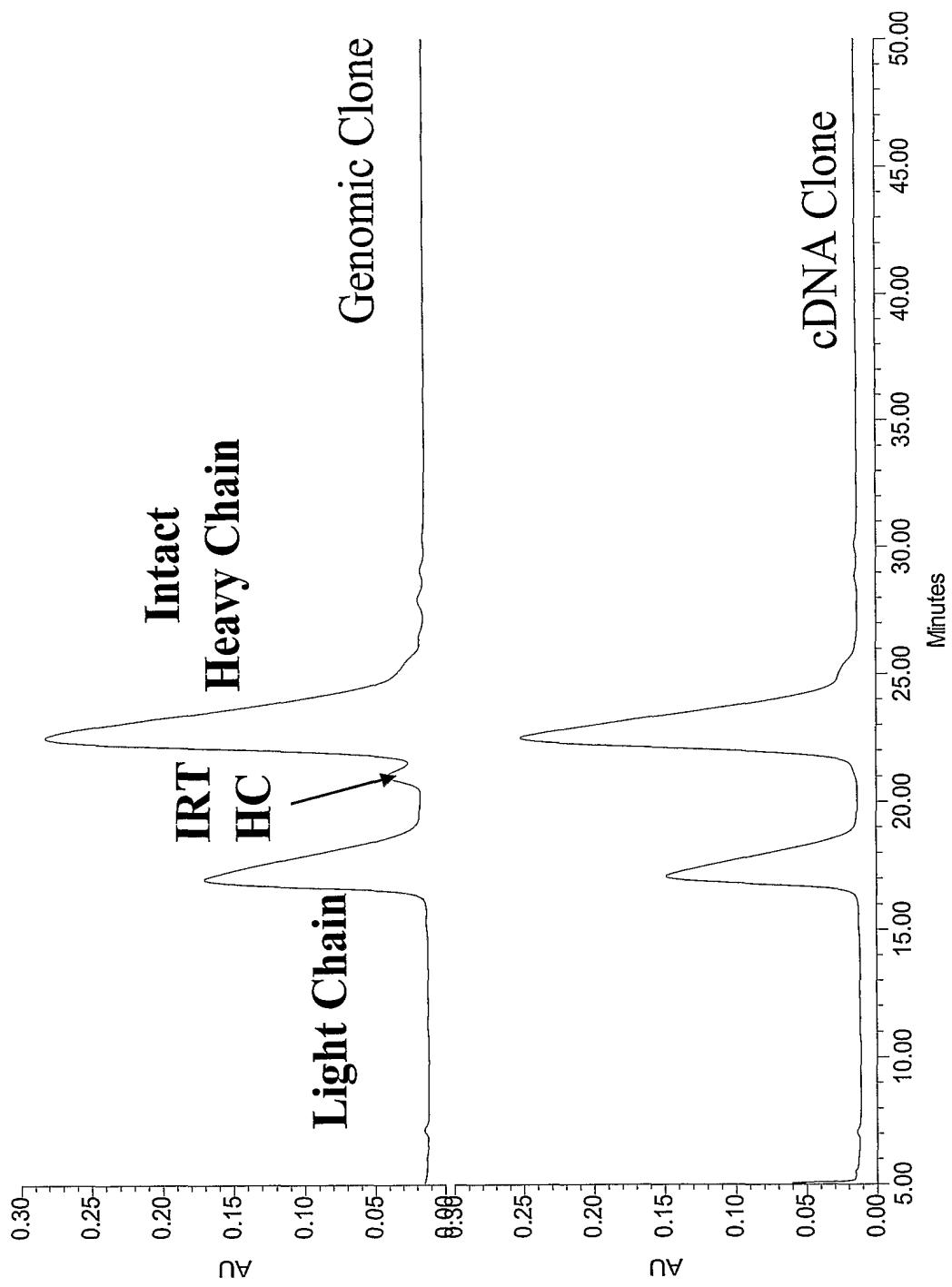


Figure 6

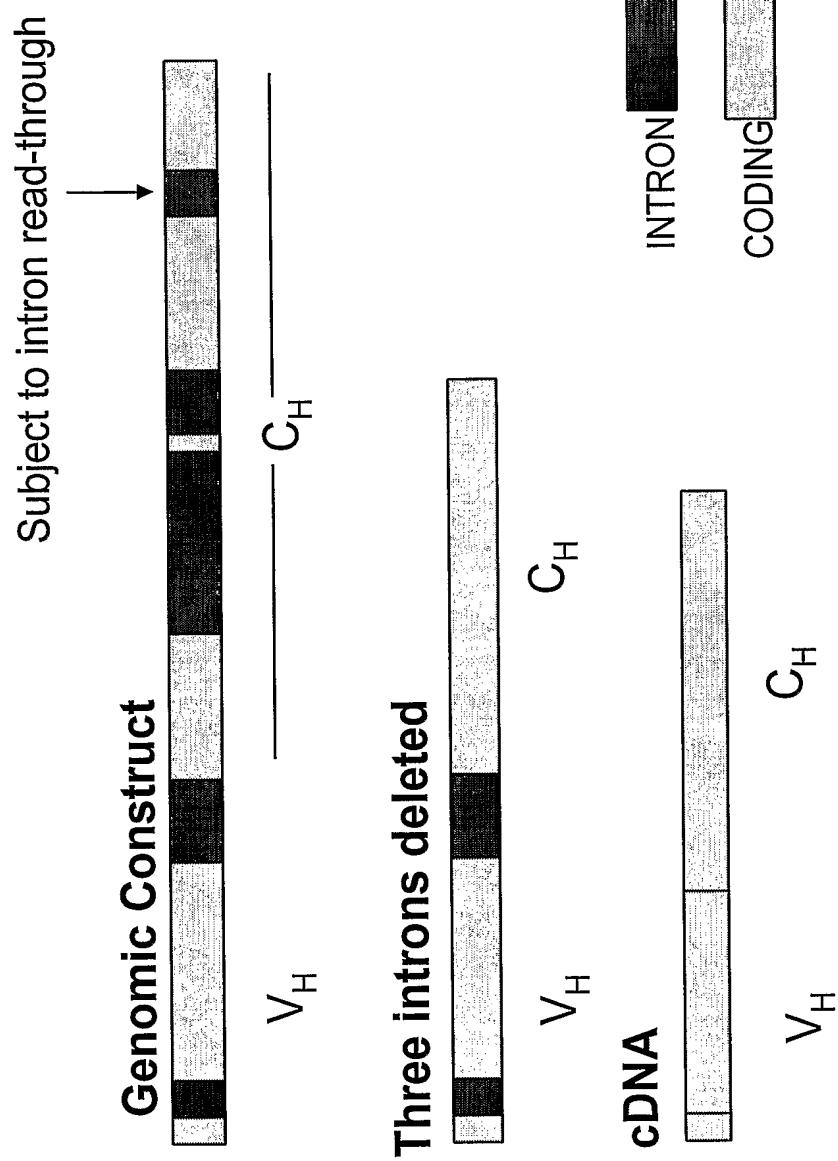


Figure 7

1 GTGAGTCCTG TCGACTCTAG AGCTTCTGG GGCAGGCCAG GCCTGACTTT GGCTGGGGC
 61 AGGGAGGGGG CTAAGGTGAC CGAGGTGGCG CCAGCCAGGC GCACACCCAA TGCCCATGAG
 121 CCCAGACACT GGACGCTGAA CCTCGCGGAC AGTTAAGAAC CCAGGGGCCT CTGCGCCCTG
 181 GGCCCAGCTC TGTCCCACAC CGCGGTCACA TGGCACCAAC TCTCTTGAG CCTCCACCAA
 A S T K
 241 GGGCCCACATCG GTCTTCCCCC TGGCACCCCTC CTCCAAGAGC ACCTCTGGGG GCACAGCGGC
 G P S V F P L A P S S K S T S G G T A A
 301 CCTGGGCTGCG CTGGTCAAGG ACTACTTCCC CGAACCGGTG ACGGTGTCGT GGAACTCAGG
 L G C L V K D Y F P E P V T V S W N S G
 361 CGCCCTGACC AGCGGCGTGC ACACCTTCCC GGCTGTCCTA CAGTCCTCAG GACTCTACTC
 A L T S G V H T F P A V L Q S S G L Y S
 421 CCTCAGCAGC GTGGTGACCG TGCCCTCCAG CAGCTTGGGC ACCCAGACCT ACATCTGCAA
 L S S V V T V P S S S L G T Q T Y I C N
 481 CGTGAATCAC AAGCCCAGCA ACACCAAGGT GGACAAGAAA GTTGGTGAGA GGCCAGCACA
 V N H K P S N T K V D K K V
 541 GGGAGGGAGG GTGTCGCTG GAAGCCAGGC TCAGCGCTCC TGCCTGGACG CATCCGGCT
 601 ATGAGTCCTC AGTCCAGGGC AGCAAGGCAG GCCCCGTCTG CCTCTTCACC CGGAGGCCTC
 661 TGCCCGCCCC ACTCATGCTC AGGGAGAGGG TCTTCTGGCT TTTTCCCCAG GCTCTGGCA
 721 GGCACAGGCT AGGTGCCCT AACCCAGGCC CTGACACCAA AGGGCAGGT GCTGGGCTCA
 781 GACCTGCCAA GAGCCATATC CGGGAGGACC CTGCCCCCTGA CCTAAGCCCA CCCAAAGGC
 841 CAAACTCTCC ACTCCCTCAG CTCGGACACC TTCTCTCCTC CCAGATTCA GTAACTCCCA
 901 ATCTTCTCTC TGCAAGAGCCC AAATCTTGAG ACAAAACTCA CACATGCCCA CGTGCCCCAG
 E P K S C D K T H T C P P C P
 961 GTAAAGCCAGC CCAGGCCCTCG CCCTCCAGCT CAAGGGCGGG AAGGTGCCCT AGAGTAGCCT
 1021 GCATCCAGGG ACAGGGCCCA GCCGGGTGCT GACACGTCCA CCTCCATCTC TTCCTCAGCA
 A
 1081 CCTGAACCTCC TGGGGGGACC GTCAGTCTTC CTCTTCCCCC CAAACCCAA GGACACCCCTC
 P E L L G G P S V F L F P P K P K D T L
 1141 ATGATCTCCC GGACCCCTGA GGTACATGC GTGGTGGTGG ACGTGAGCCA CGAACACCT
 M I S R T P E V T C V V V D V S H E D P
 1201 GAGGTCAAGT TCAACTGGTA CGTGGACGGC GTGGAGGTGC ATAATGCCAA GACAAAGCCG
 E V K F N W Y V D G V E V H N A K T K P
 1261 CGGGAGGGAGC AGTACAAACAG CACGTACCGT GTGGTCAGCG TCCTCACCGT CCTGCACCA
 R E E Q Y N S T Y R V V S V L T V L H Q
 1321 GACTGGCTGA ATGGCAAGGA GTACAAGTGC AAGGTCTCCA ACAAAAGCCCT CCCAGCCCC
 D W L N G K E Y K C K V S N K A L P A P
 1381 ATCGAGAAAA CCATCTCCAA AGCCAAAGGT GGGACCCGTG GGGTGCAGG GCCACATGGA
 I E K T I S K A K
 1441 CAGAGGCCGG CTCGGCCAC CCTCTGCCCT GAGAGTGACC GCTGTACCAA CCTCTGTCCC
 1501 TACAGGGCAG CCCCAGAAC CACAGGTGTA CACCCCTGCC CCATCCCGGG AGGAGATGAC
 G Q P R E P Q V Y T L P P S R E E M T
 1561 CAAGAACCAAG GTCAGCCTGA CCTGCCTGGT CAAAGGCTTC TATCCAGCG ACATGCCGT
 K N Q V S L T C L V K G F Y P S D I A V
 1621 GGAGTGGGAG AGCAATGGGC AGCCGGAGAA CAACTACAAG ACCACGCCCTC CCGTGCTGGA
 E W E S N G Q P E N N Y K T T P P V L D
 1681 CTCCGACGGC TCCTTCTTCC TCTATAGCAA GCTCACCGTG GACAAGAGCA GGTGGCAGCA
 S D G S F F L Y S K L T V D K S R W Q Q
 1741 GGGGAACGTC TTCTCATGCT CCGTGATGCA TGAGGGCTCTG CACAACCACT ACACGCAGAA
 G N V F S C S V M H E A L H N H Y T Q K
 1801 GAGCCCTCTCC CTGTCCCCGG GTAAATGA (SEQ ID NO:1)
 S L S L S P G K (SEQ ID NO:2)

Figure 8

Figure 9

10/11

GTGAGTCCTGGACTCTAGAGCTTCTGGGCCAGGCCACACCAATGCCATTGAGCCCAGCAGGTGGCTGGCTGGGGCAGGGCTGGCTGGGGCT
 AAGGTGACGCCAGGTGGCCAGGCCACACCAATGCCATTGAGCCCAGCAGGTGGCTGGCTGGGGCTGGCTGGGGCTGGCTGGGGCTGGCTGGGGCT
 CGGGACAGTTAAGAACCCAGGGGCCTCTGGGCCAGGCCACACCAATGCCATTGAGCCCAGCAGGTGGCTGGCTGGGGCTGGCTGGGGCTGGCTGG
 CACCTCTCTTGCAAGCCTCCACCAAGGGCCATCGGTCTCCCCCTGGCACCCCTCCTCCAAAGAGCACCTCTGG
 GGGCACAGGGCCCTGGCTGGCTGGCTGGCTGGGGCTGGCTGGGGCTGGCTGGGGCTGGCTGGGGCTGGCTGGGGCTGGCTGGGGCTGGCTGG
 CGCCCTGACAGGGGGTGCACACCCTCCGGCTACAGTCCCTCAGGACTCTACTCCCTCAGCAGCGT
 GGTGACCGTGCCTCCAGCAGCTGGCACCCAGACCTACATCTGCACAGTGAATCACAAAGGCCAGCAACAC
 CAAGGTGGACAAGAAAGTTGGTAGAGGGCTATGGCAGTCCCCAGTCCAGGGCAGCAAGGCAGGGCCCTCTGGCTCTGGCA
 CCCGGAGGCCTCTGGCCGCCACTCATGGTAGAGGGTCTCAGGGAGGGTCAACATGGCTTTGGCTTTGGCTCTGGCTCTGGCA
 GGCACAGGCTAGGTGCCCCATAACCCAGGCCCTGACACACAAGGGGAGGGCTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGG
 GCCATATCCCCGGAGGACCCCTGGCCCTGACCTTAAGGCCACCCAAAGGCCAAACTCTCCACTCCCTCAGCTCG
 GACACCTCTCTCCCTCCAGATTCCAGTAACTCCCTCAGCTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGG
 CTCACACATGCCACCCAGGGTGGCCAGGTAAGCCAGGCCAGGGTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGG
 CTAGAGTAGCCTGCATCCAGGGACAGGCCAGGCCAGGGTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGGCTGG
 CCTGGAACTCCTGGGGACCCGTCAAGTCCTTCCCTCTCCAGGAAAGGCCAGCCCTGAGGTCAAGTCAACTGGTACGGT
 ACCCCTGAGGTACATGGTGGTGGGGACGGTCAAGTCAACTGGTACGGTCAAGTCAACTGGTACGGT
 GACGGCGTGGAGGTGCAATAATGCCAAGACAAAGCCGGAGGGAGGAGCTGAACACAGCACGGTACACAGCAGGTGCTGGCTGGCTGGCT
 AGCGTCCCTCACCGTCCCTGCACGGACTGGCTGAATGGCAAGGGAGTACAAGTGAAGGTCTCCAAACAAAGCC
 CTCCCAAGCCCCATCGAGAAAACCATTCCAAAGCCAAAGGGCAGCCCCGAGAACCAAGGGCTTCTATCCCAGC
 CCCCATCCCCGGAGGAGATGACCAAGAACCGGTCAAGCCGGAGAACAAACTACAAGAACGCGCTCCGGTGGCTGGAC
 GACATGCCCGTGGAGTGGAGAACCAAGGGCAGCCGGAGAACAAACTACAAGAACGCGCTCCGGTGGCTGGAC
 TCCGACGGCTCCTTCTTCCCTATAGCAAGGCTCACCGTGGACAAAGAGCAGGTGGCAGGGAAACGTCTTC
 TCATGCTCCCGTGTGATGCATGAGGCTCTGGCTCGCACAAACCAACTACACCGAGAACAGGCTTCTCCCCGTGGTAAA
 TGA (SEQ ID NO: 5)

Figure 10

GTGAGTCCTCGACTCTAGAGCTTCTGGGCAGGCCAGGCCAGGCCACACCCAAATGCCCATGAGCCAGACACTGGACGCTGAACCTCGCGG
GGTGAACGCCAGGTGGCCAGGCCAGGCCACACCCAAATGCCCATGAGCCAGACACTGGACGCTGAACCTCGCGG
ACAGTTAAGAACCCAGGGCCCTCTGGCCCTGGCCCTGGCCCTGGCCAGCTCTGGCCACACCCCAATGCCCATGAGCCAGACACTGGACGCTGAACCTCGCGG
TCTTGAGCCTCCACCACAGGGCCATCGGTCTTCCCCCTGCGGCCCTGCTCCAGGAGCACCTCCGAGAGCACAG
CGGGCCCTGGGCTGCCCTGGTCAAGGACTACTCCCCGAACCCGGTGACCGTGTGGAAACTCAGGGCCCTGACC
AGCGGCGTGCACACCTTCCCGCTGTCTACAGTCCCTCAGGACTCTACTCCCCACAGCCAGAACACACCAAGGTGGACAAGA
CTCCAGCAGCTTGGCACGGCACGAAAGACCTACACCTGCAATGTAGATCACAGCCAGAACACACCAAGGTGGACAAGA
GAGTTGAGTCCAAATATGGTCCCCCATGCCACCTGAGTTCTGGGGACCATCAGTCTTC
CTGTTCCCCCAAACCCAAAGGACACTCTCATGATCTCCGGACCCCTGAGGTCACTGGTGGCTGGGTGGACGT
GAGCCAGGAGACCCCCAGGGTCCAGTCAACTGGTAACCTGGTCAACCTGGTCAACAGCAGTCAACAGCACGTACCGTGTGGTCAACCGTCTCACCCTGCACCAAGGACTGGCTGAAC
GGCAAGGAGTACAAGTGCAAGGTCTCCAACAAAGGCCCTCCCATCGAGAAAACCATCTCCAAAGCCAA
AGGGCAGCCCCGAGAGCCACAGGTGTACACCCCTGCCCATCCCCATCCAGGAGGAGATGACCAAGAACCCAGGTAGCC
TGACCTGCCTGGTCAAAGGCTTCTACCCAGGGACATCGCCGCTCCCTGGAGTGGAGGAAATGGCAGCCGGAGAAC
AACTACAAGAACACGGCTCCCGTGGACTCCGACGGCTCCCTACAGCAGGGCTAACCCGTGGACAA
GAGCAGGTGGCAGGAGGGAAATGTCTCATGCTCCGTGATGCATGAGGCTCTGCACAAACCTACACACAGA
AGAGCCTCTCCCTGTCTGGTAAATGA (SEQ ID NO: 6)

Figure 11

INTERNATIONAL SEARCH REPORT

International application No
PCT/US2005/036247

| | | | | | |
|--|----------------|------------|-----------|----------|----------|
| A. CLASSIFICATION OF SUBJECT MATTER | INV. C12N15/09 | C12N15/13 | C12N15/67 | C12N5/10 | C12Q1/68 |
| | C07K16/00 | A61K39/395 | | | |

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
C12N C12Q C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, BIOSIS, Sequence Search

C. DOCUMENTS CONSIDERED TO BE RELEVANT

| Category* | Citation of document, with indication, where appropriate, of the relevant passages | Relevant to claim No. |
|-----------|---|-----------------------|
| X | <p>CHOI T ET AL: "A GENERIC INTRON INCREASES GENE EXPRESSION IN TRANSGENIC MICE" MOLECULAR AND CELLULAR BIOLOGY, vol. 11, no. 6, 1991, pages 3070-3074, XP007900213 ISSN: 0270-7306 abstract</p> <p>-----</p> <p style="text-align: center;">-/--</p> | 1-53 |

Further documents are listed in the continuation of Box C.

See patent family annex.

* Special categories of cited documents :

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Name and mailing address of the ISA/
European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl
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